

# Structure and Measurement of the brain lecture notes

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2009/2010

Based on slides from Flavia Filimon, 2008

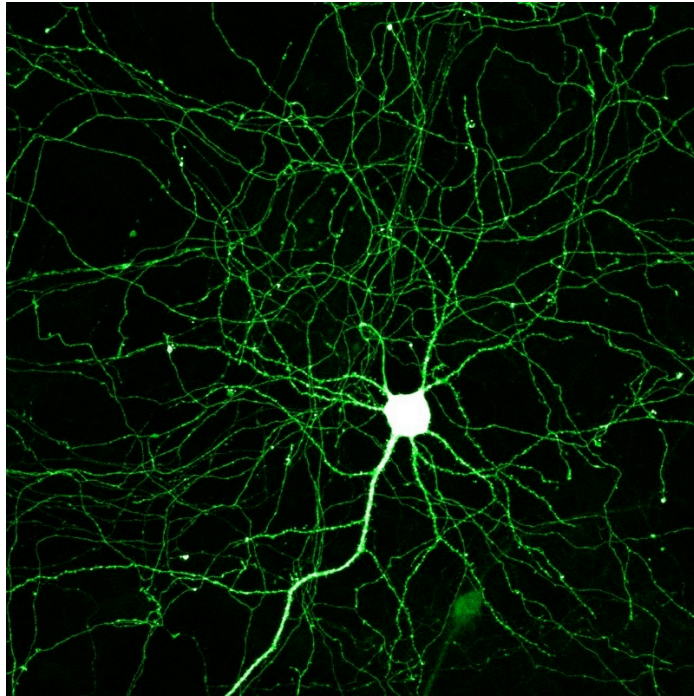
# Neurons and Models

## Lecture I

# Topics

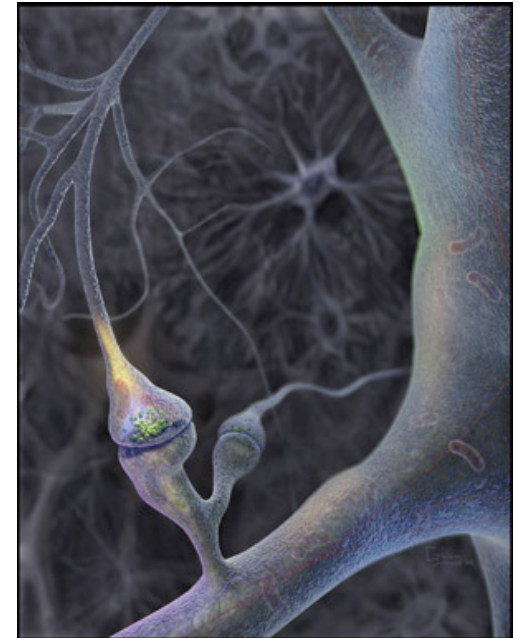
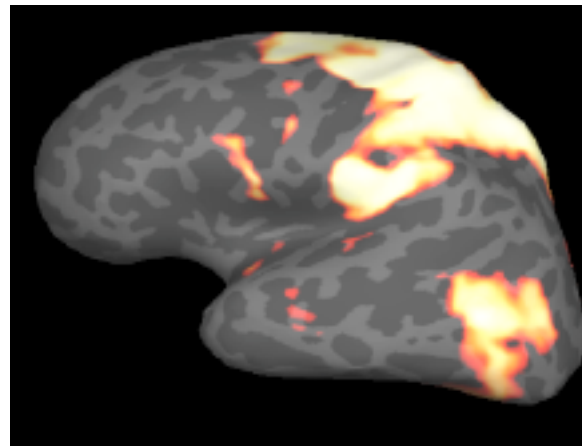
- Membrane (Nernst) Potential
- Action potential/Voltage-gated channels
- Post-synaptic potentials, ligand gated channels
- Dendritic propagation equivalent circuits
- NMDA channels and synaptic plasticity
- Spike timing dependent plasticity (STDP)

# How does the brain work?

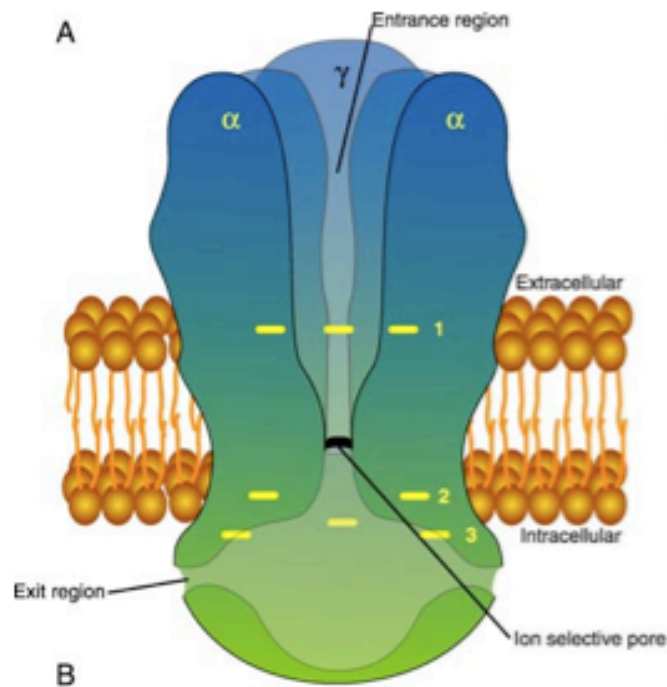


- 100 billion neurons in the human brain
- $10^{14}$  synapses (1000-5000 per neuron)

from molecular  
level  
to systems level



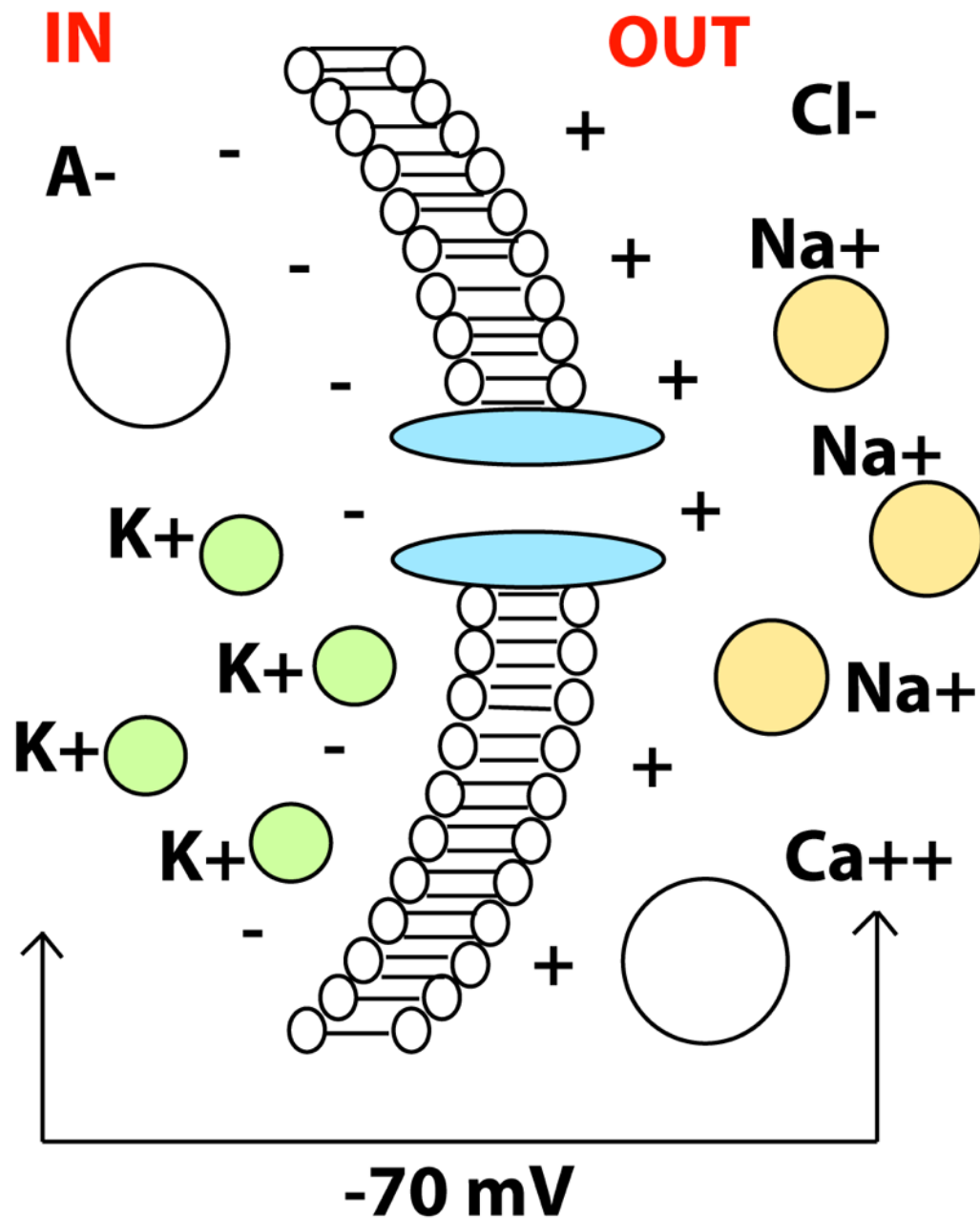
# Ion channels



- resting (permanently open at rest)
- gated (require ligand, voltage, or mechanical stretch, to open)

# Membrane Potential

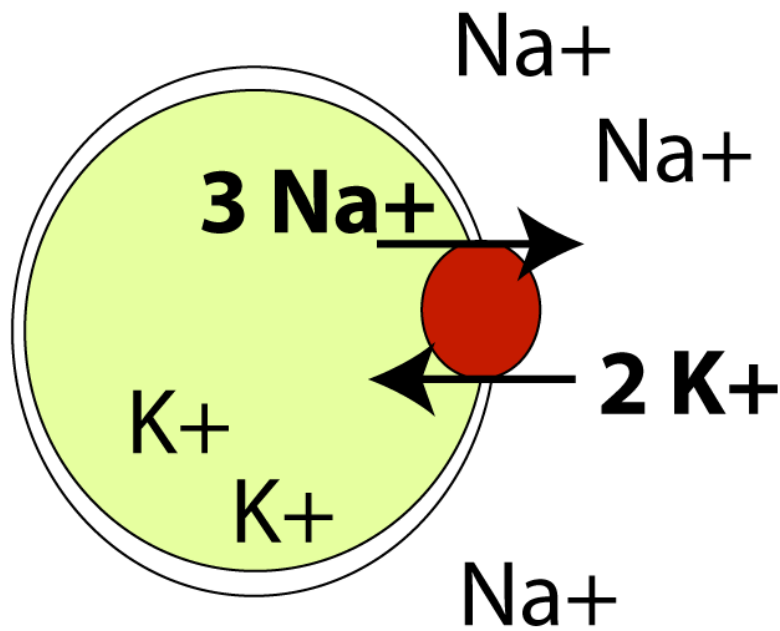
- $V_m$  (membrane pot.) due to \*resting\* channels
- = voltage difference across the membrane
- I. different ions have different concentration gradients across the membrane
- ion species:  $K^+$ ,  $Na^+$ ,  $Cl^-$ ,  $Ca^{++}$
- II. membrane is semi-permeable - most resting channels are  $K^+$  (leaky) channels



## Membrane Potential ( $V_m$ )

- $\sim -70$  mV (depends on cell type)
- semi-permeable membrane:  $K^+$
- differential concentration gradients of  $K^+$ ,  $Na^+$ ,  $Cl^-$ ,  $Ca^{++}$

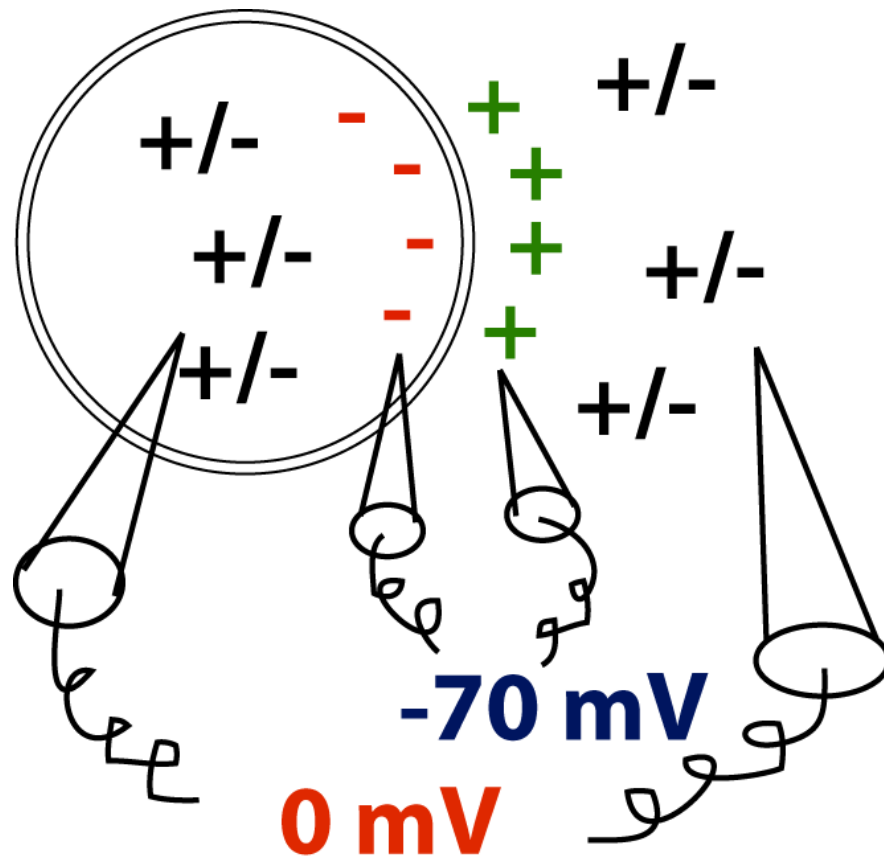
# Na<sup>+</sup> - K<sup>+</sup> pump



- 3 Na<sup>+</sup> out, 2 K<sup>+</sup> in
- moves ions against their concentration gradient
- re-establishes concentration gradients



note:

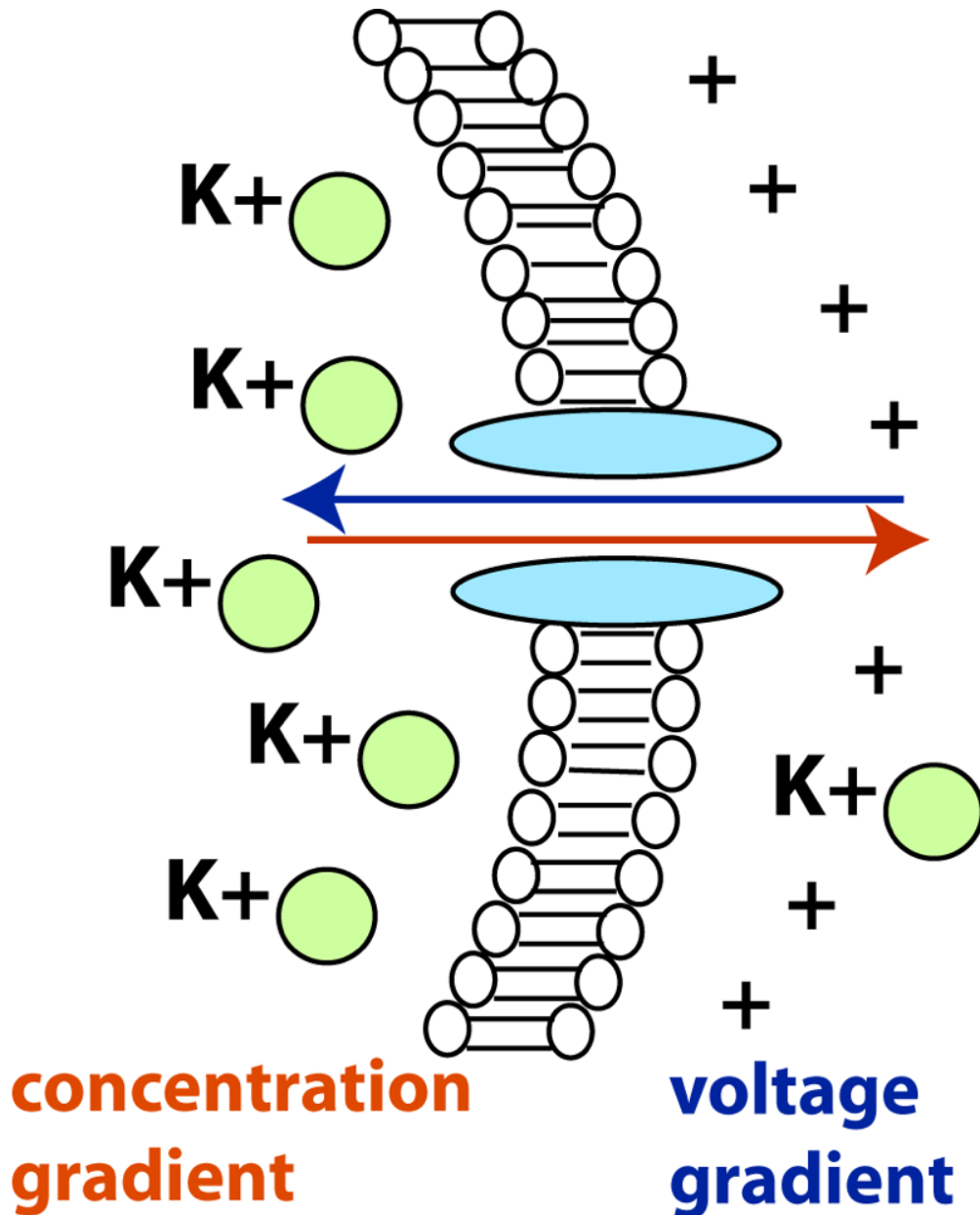


- voltage & concentration difference only immediately across membrane

# Purpose of resting potential?

- signaling is a brief deviation from the resting potential;
- to signal information, must have a baseline/ resting state so incoming information isn't drowned in noise

# Nernst Potential



- equilibrium potential for one ion
- = reversal potential
- when concentration gradient force balances out electrical force

# Nernst values for different ions (in mammalian neurons)

	$[\text{ion}]_i$ (mM)	$[\text{ion}]_o$ (mM)	$E_{\text{ion}}$ (mV)
K <sup>+</sup>	135	3	-102
Na <sup>+</sup>	18	150	+56
Cl <sup>-</sup>	7	120	-76
Ca <sup>++</sup>	0.1 $\mu\text{M}$	1.2	+125

# Nernst potential

- NERNST EQUATION - target potential for one ion that must be distributed both inside and outside the cell
- reversal potential:  $V_m$  above or below Nernst: ion current reverses direction
- equilibrium potential = Nernst potential if channel permeable to only 1 ion (note: a channel can also have a Nernst potential)

# Nernst questions

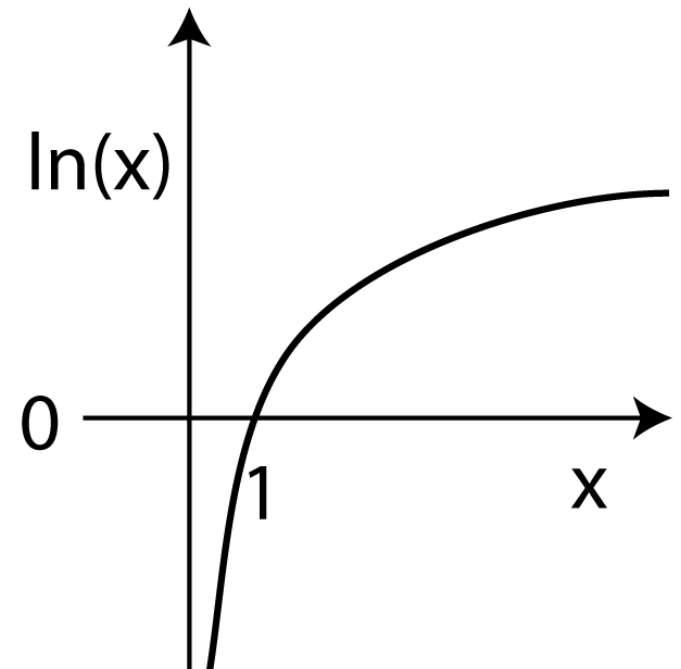
- Q: what happens to  $K^+$  if  $V_m$  is lowered to  $-130$  mV? What about if it is raised to  $-50$  mV?
  - $\rightarrow K^+$  moves in; 2)  $K^+$  leaves cell
- Q: What happens to  $Na^+$  if channels are closed, and membrane potential is raised to  $-40$  mV? (Nothing: channels are closed, can't get in). How about: raising  $V_m$  to  $+65$  mV?
  - $\rightarrow$  if channels open,  $Na^+$  will leave the cell

# Nernst Equation

- allows to calculate Nernst potential for one ion

$$E_{\text{ion}} = RT/zF * \ln([\text{ion}]_o/[\text{ion}]_i)$$

- $z$  = valence (+/- 1 or for  $\text{Ca}^{++}$ : +2)
- $\ln(>1)$  = +ve number;  $\ln(<1)$  = -ve number
- $\ln(1) = 0$  --> Nernst will be zero.



# Equilibrium potential continued

- +ve ion more concentrated outside  $\rightarrow$  +ve  $E_{ion}$
- +ve ion more concentrated inside  $\rightarrow$  -ve  $E_{ion}$
- -ve ion more concentrated outside  $\rightarrow$  -ve  $E_{ion}$
- -ve ion more concentrated inside  $\rightarrow$  +ve  $E_{ion}$
- Question: Suppose you have a species of ion called Flavium which is +ve, and has a -ve Nernst potential. Are there more Flavium ions inside or outside the cell?
- ( $\rightarrow$  inside)



## GOLDMAN EQUATION

$$V_m = \frac{RT}{F} \cdot \ln \left[ \frac{(p_K[K^+]_o + p_{Na}[Na^+]_o + p_{Cl}[Cl^-]_i)}{(p_K[K^+]_i + p_{Na}[Na^+]_i + p_{Cl}[Cl^-]_o)} \right]$$

- calculates  $V_m$  for multiple ions
- permeability of membrane to ions and concentration (inside vs. outside) of ions
- $K^+$ ,  $Cl^-$ , and  $Na^+$  all contribute to the resting membrane potential; but membrane more permeable to  $K^+$

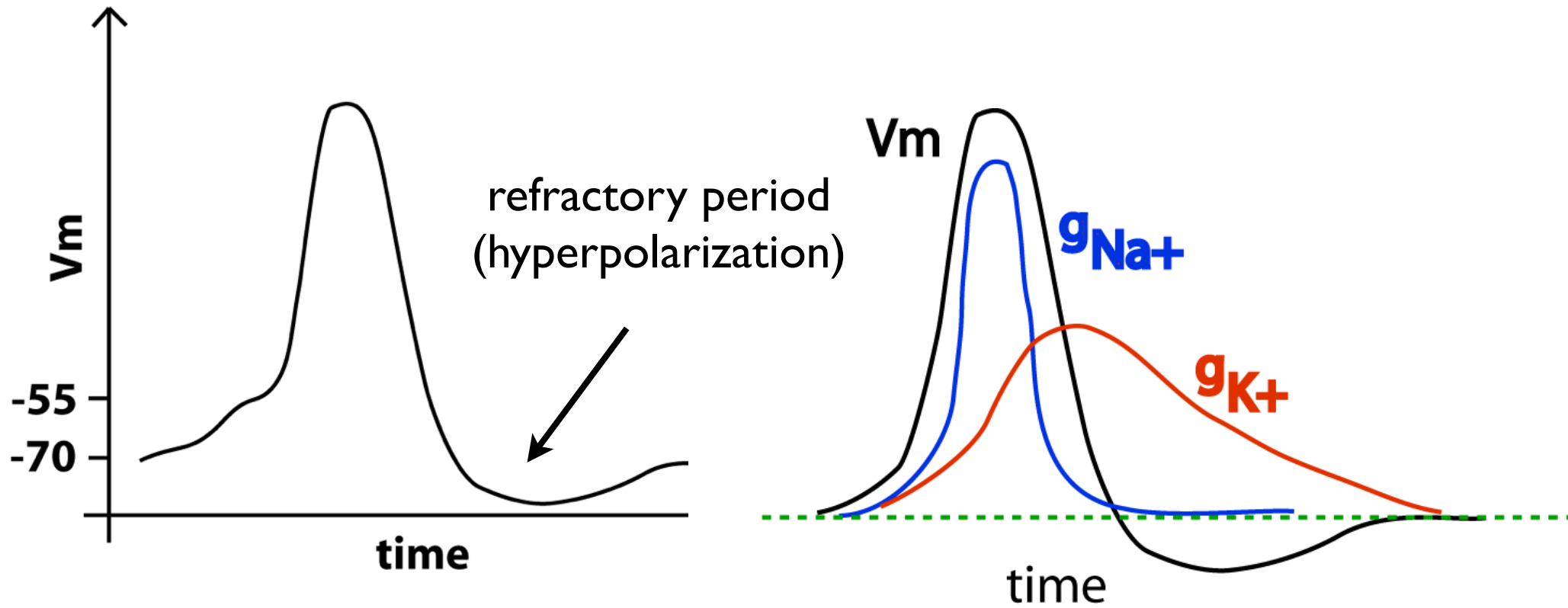
# Questions

- What happens if you tear a hole in the cell membrane?
  - $V_m$  goes to zero, cell dies (after spiking a lot due to depolarization)
- What happens if you add  $K^+$  ( $K^+Cl^-$ ) outside the cell at rest?
  - $K^+$  enters cell, depolarizes it

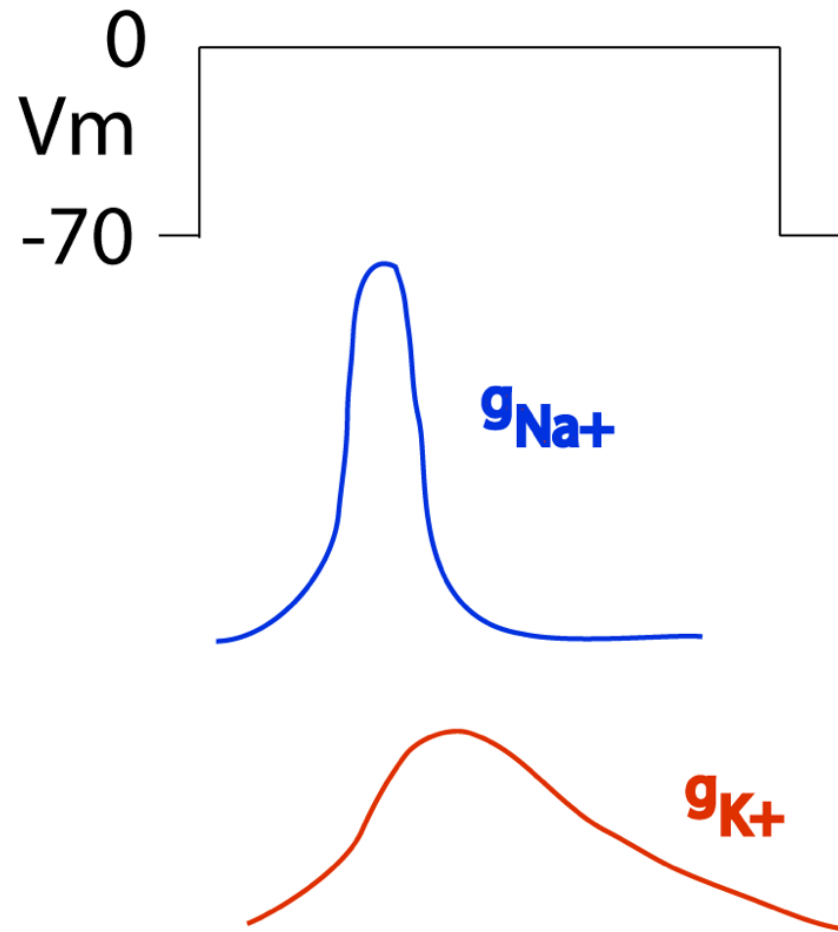
# Action Potential

- Purpose: long-distance communication; e.g. photoreceptor cells in retina don't need to spike, b/c other cells are close-by
- depends on voltage-gated  $\text{Na}^+$  and  $\text{K}^+$  channels
- Hodgkin-Huxley equation

# Action Potential



# Voltage-gated $\text{Na}^+$ and $\text{K}^+$ conductances

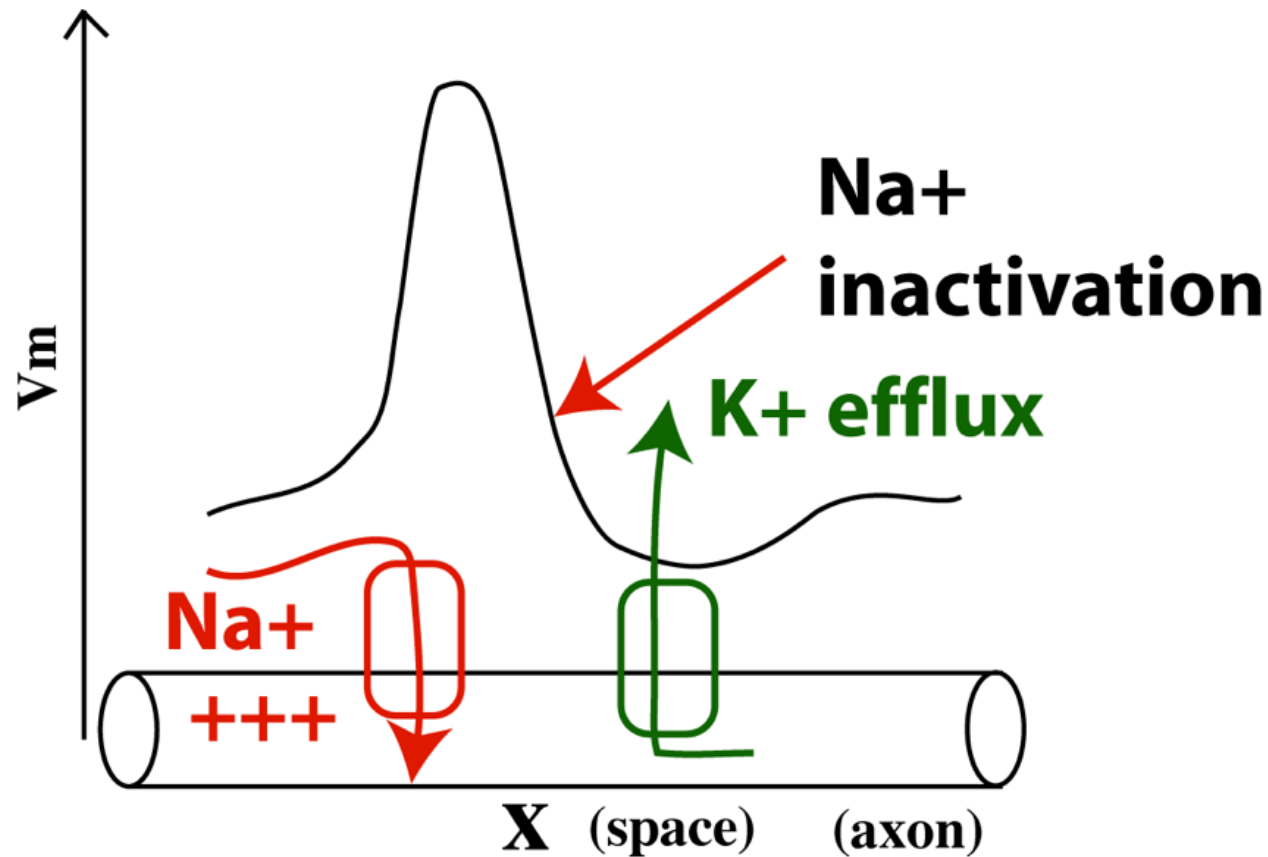


- $\text{Na}^+$ : fast, transient, inactivating
- $\text{K}^+$ : slow/delayed, long-lasting, non-inactivating

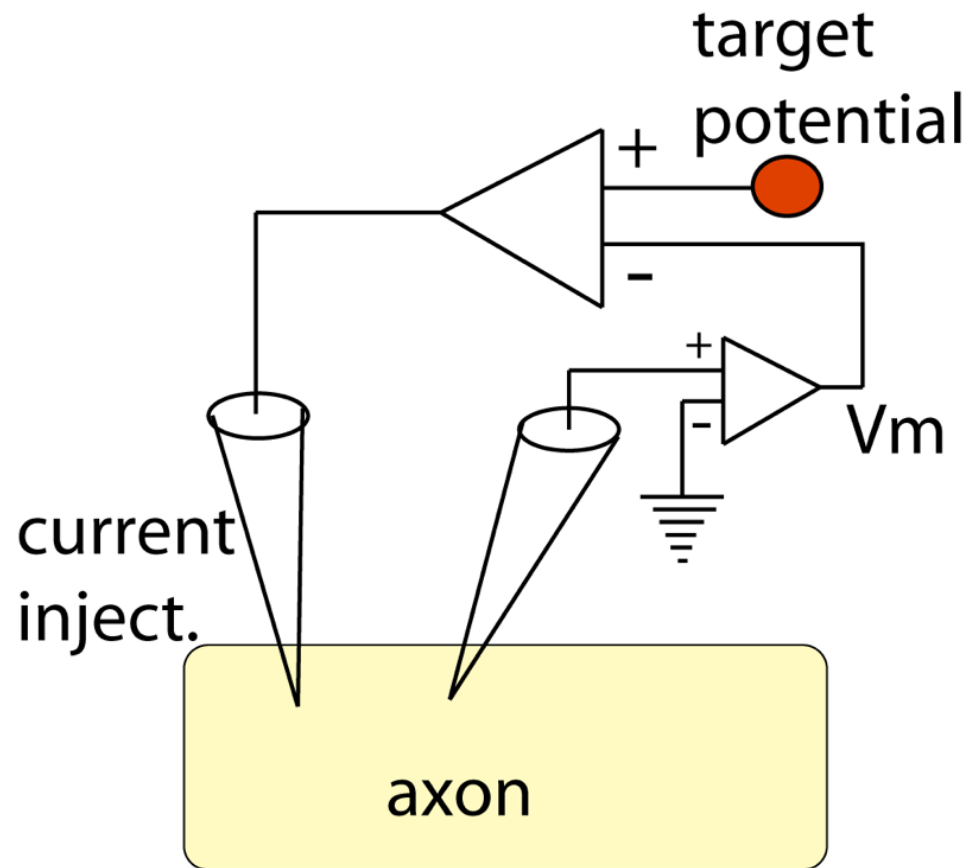
# Action Potential

- fast voltage-gated inward  $\text{Na}^+$  current that inactivates: transient
- slow long-lasting voltage-gated outward  $\text{K}^+$  current that does not inactivate, only deactivates: sustained
- purpose of  $\text{Na}^+$  inactivation: prevent reverberation; cell can't spike during absolute refractory period no matter what the voltage - not due to negative voltage, but due to inactivation of  $\text{Na}^+$  channels

# Hyperpolarization is caused by $K^+$ efflux and $Na^+$ inactivation



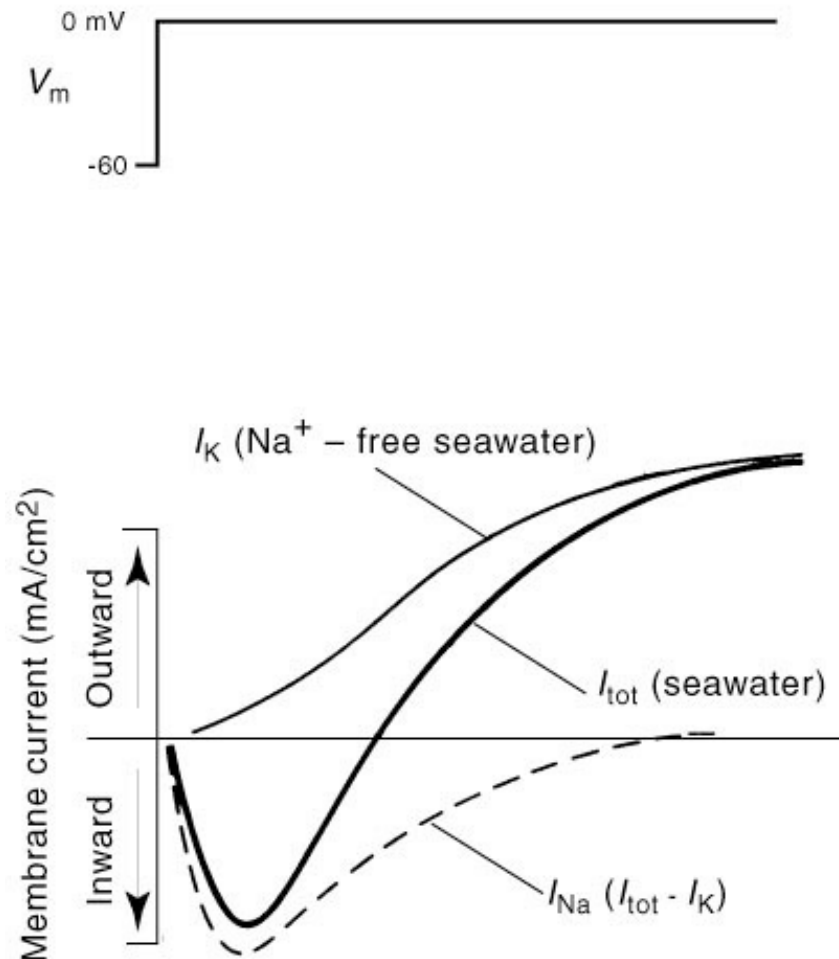
# Voltage clamp



- two electrodes: voltage electrode + current electrode
- compare desired  $V_m$  to actual  $V_m$ , inject +ve or -ve current



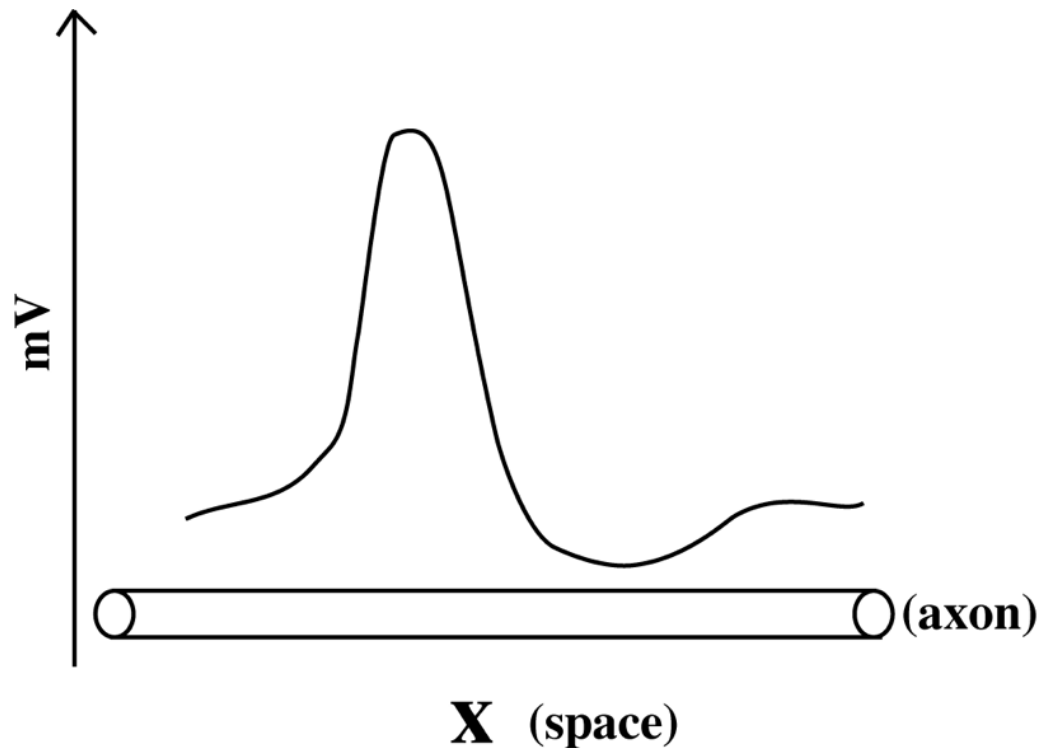
# Characterizing time course and amplitude of ionic currents during Action Potential



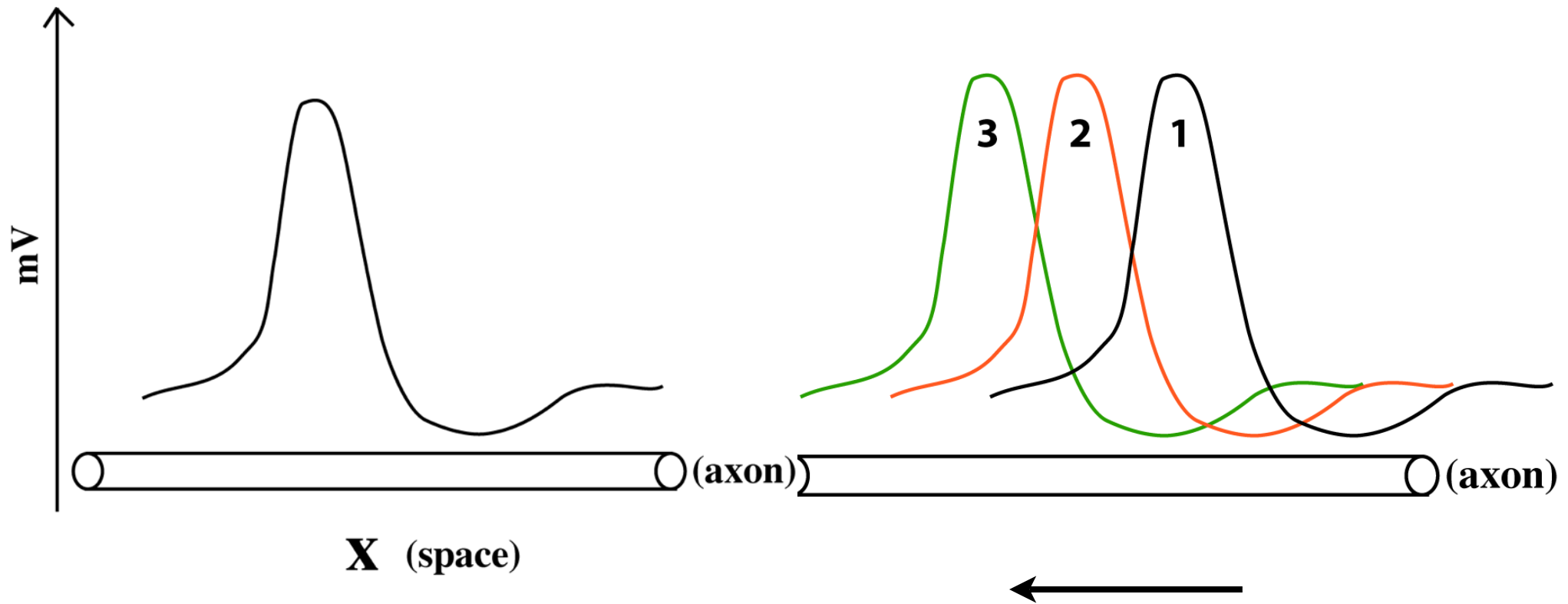
- voltage-clamp technique and selective removal of ions allows us to determine which ionic currents contribute to the AP (action pot.)

From: Fundamental Neuroscience, Squire et al. 2003

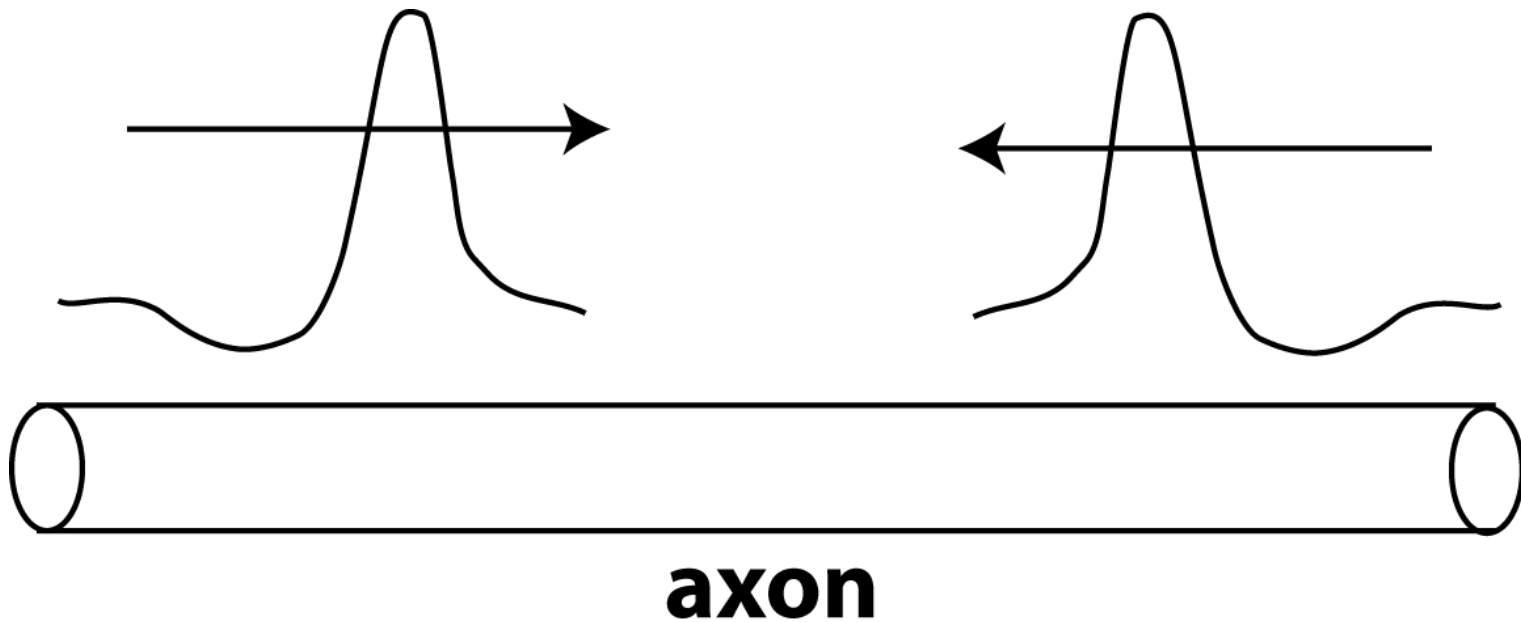
# Which way is this AP traveling?



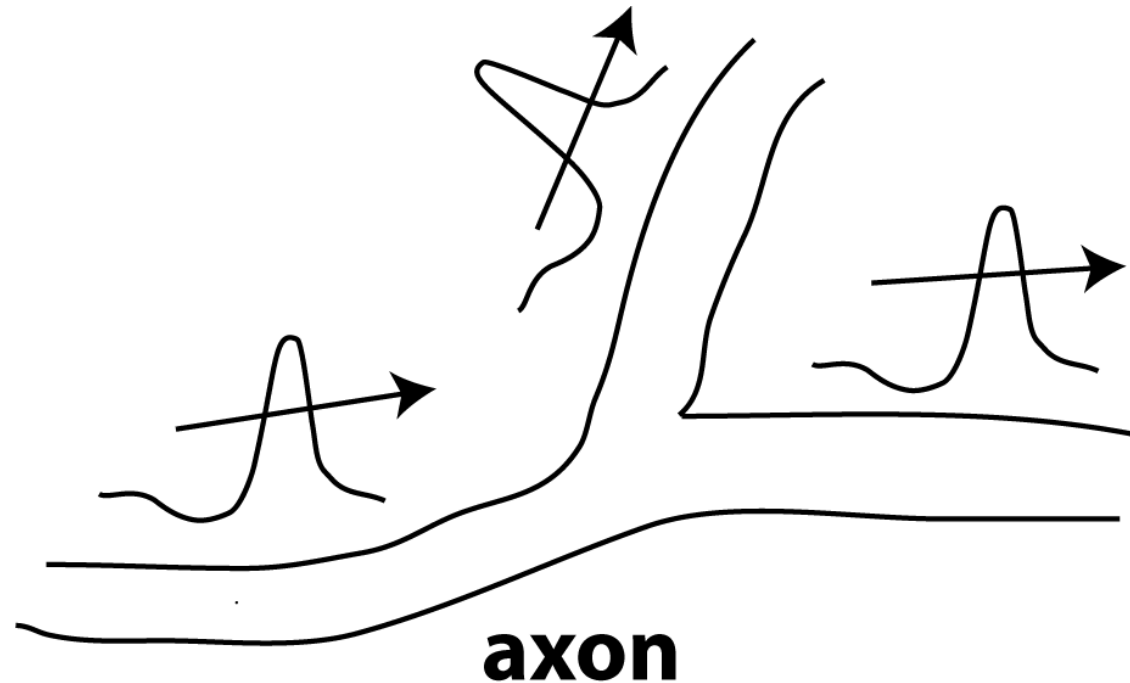
# Which way is this AP traveling?



# What happens when two APs collide?



# What happens to AP when axon splits in two?

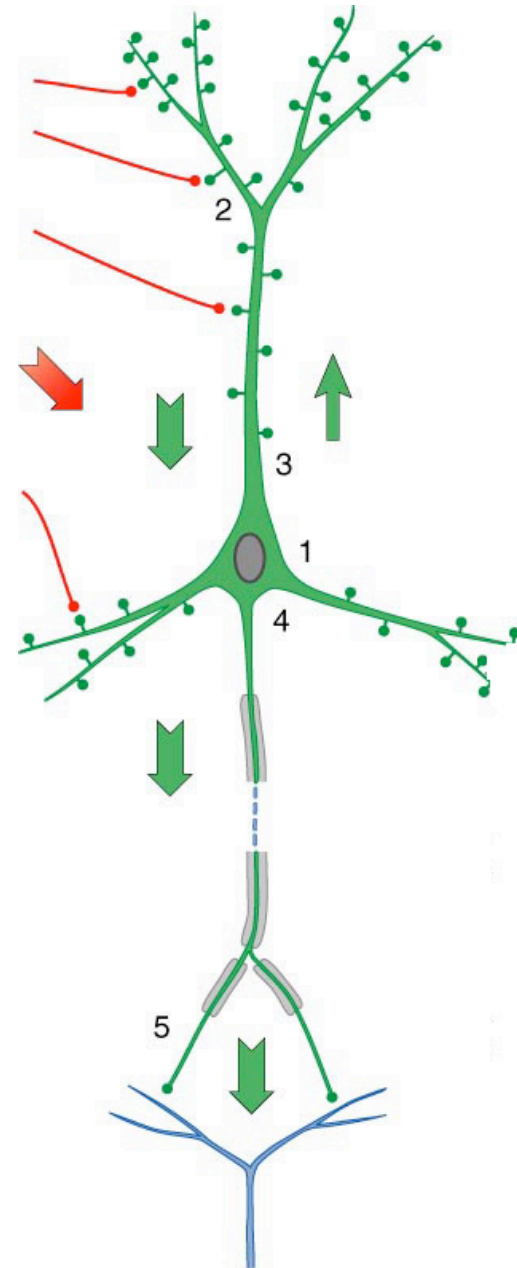


# AP amplitude does not halve

- Action Potentials are actively regenerated; i.e. same amplitude; they're "all or none" - can't have just  $1/2$  an action potential
- therefore: if an AP hits a branch in an axon, it will either die, or go down each branch with the same amplitude; it won't halve. It might die down one branch rather than the other, but it won't halve its amplitude
- contrast with "electrotonic" or "graded" potentials (passively spread).

# Electrotonic Potentials/ graded potentials

- passively spreading electric current
- (as opposed to actively *propagated* action potentials)
- usually from dendritic inputs; or current injection via electrode



# Basic concepts

- $R$  = resistance (difficulty of spreading; e.g. Library Walk)
- $I$  = current (amount of flow) ( $I = Q/t$ )
- $V$  = voltage (e.g. “water pressure”)
- $C$  = capacitance (how much charge you can hold);  $C \propto \text{area/distance betw. plates}$  (e.g. 5 nm)
- $g$  = conductance =  $1/R$
- $Q$  = charge =  $C*V$

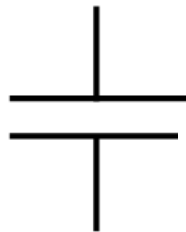


# Symbols

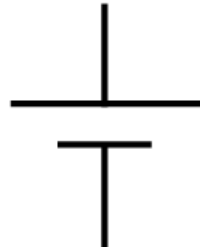
- resistor



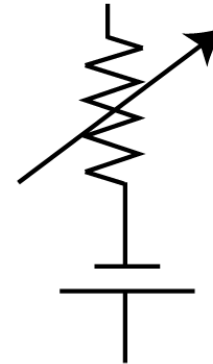
- capacitor



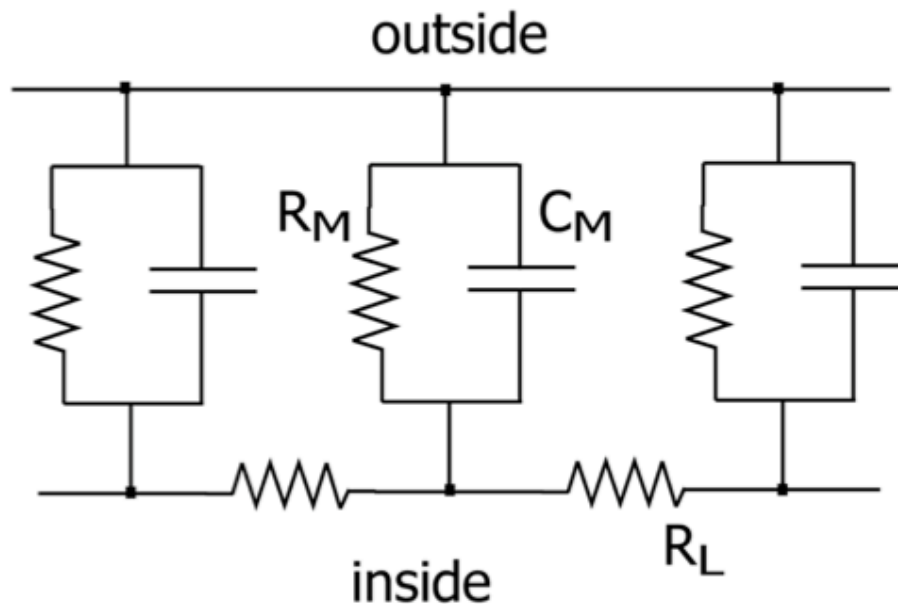
- battery



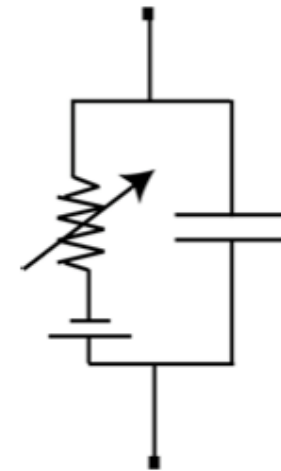
- Nernst potential across channel



# Equivalent Electrical Model of Dendrite

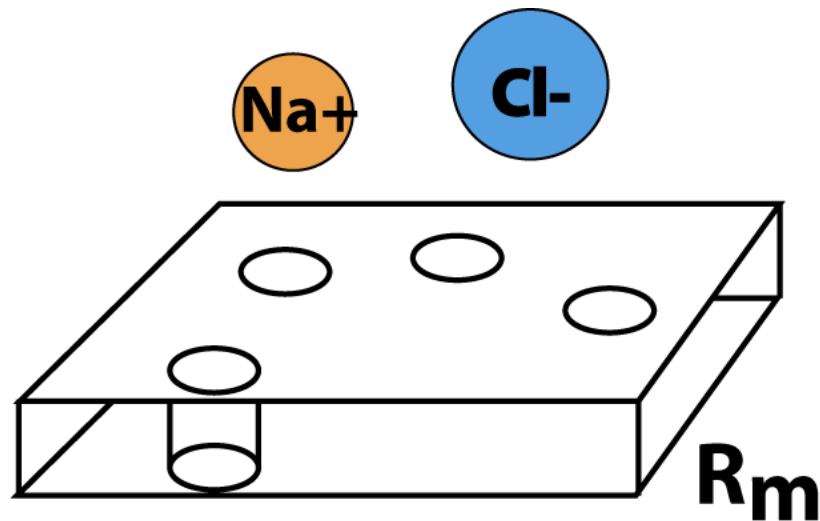


In membrane,  $C_m$  and  $R_m$  are in parallel;  $R_L$  are in series;  $R_L$  is much larger than  $R_m$

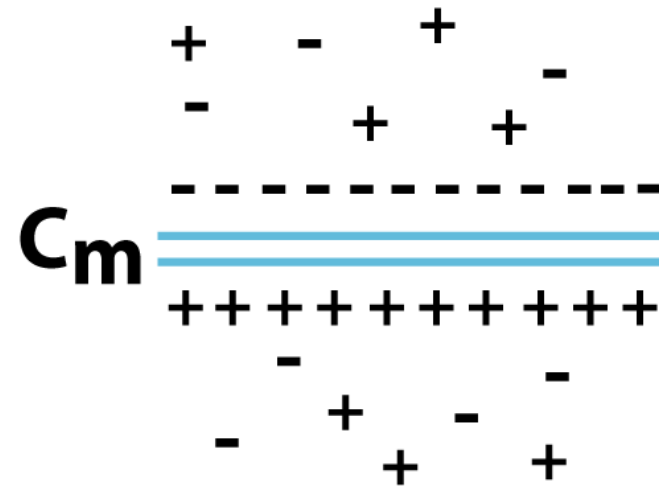


Patch of membrane with Nernst potential across channel (serves as battery)

# $R_m$ and $C_m$

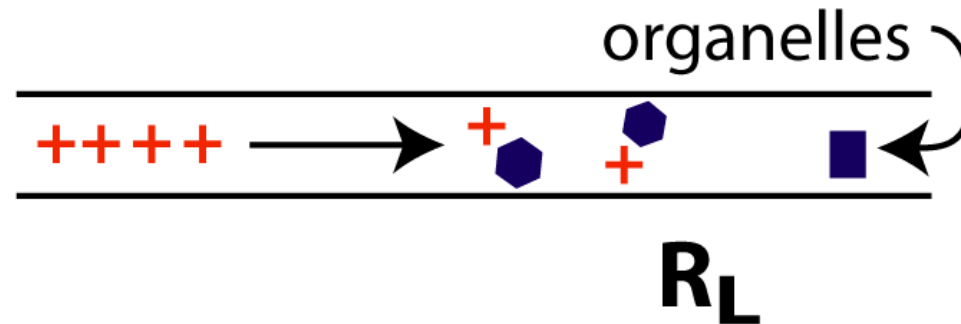


membrane has  
resistance ( $R_m$ )



membrane has  
capacitance ( $C_m$ )

$R_L$

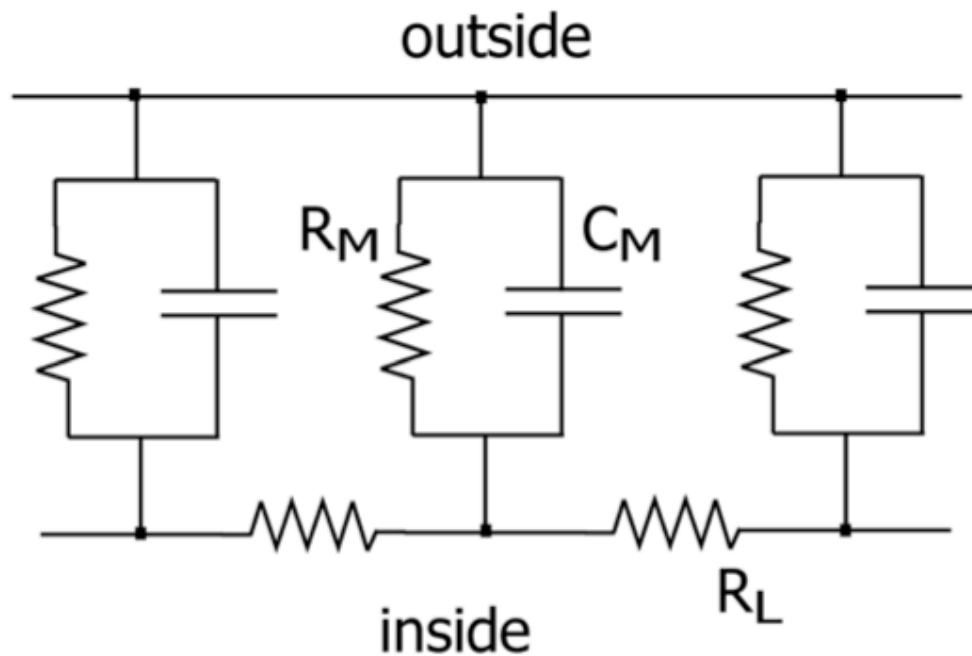


- axons/dendrites have internal/axial/longitudinal resistance ( $R_L$ )
- NOTE: outside resistance negligible (zero)

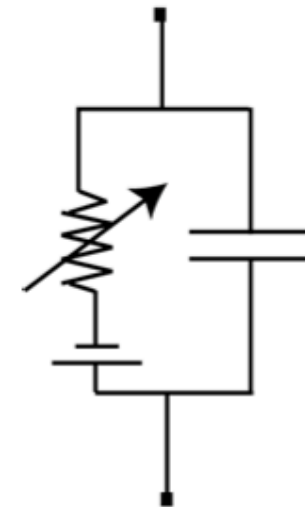
# Laws

- uncharged capacitor = zero resistance
- charged capacitor = infinite resistance
- it takes time to charge a capacitor
- current follows the path of least resistance

# Equivalent Electrical Model of Dendrite

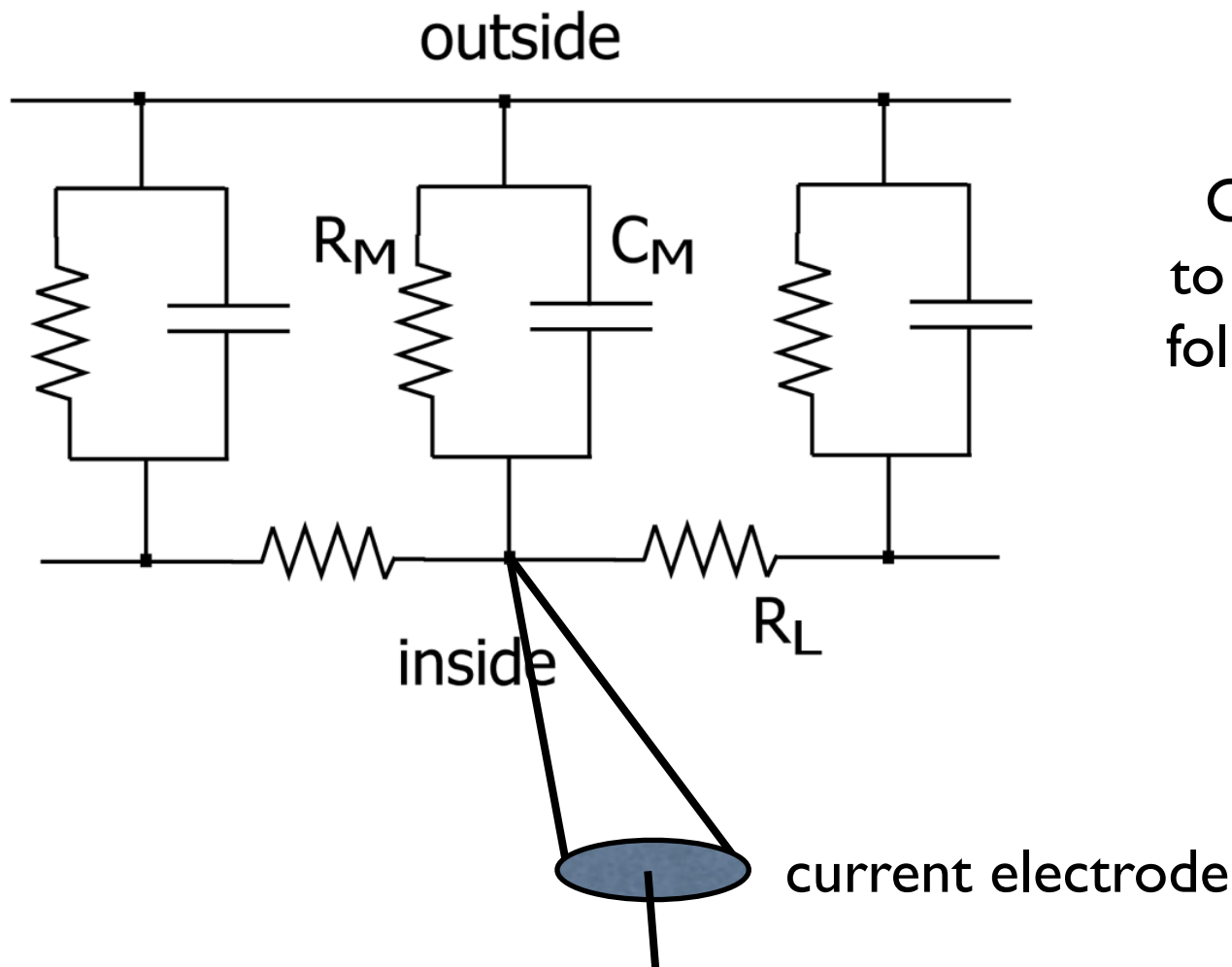


In membrane,  $C_m$  and  $R_m$  are  
in parallel;  $R_L$  are in series  
 $R_L$  is much ~~larger~~ than  $R_m$   
smaller!



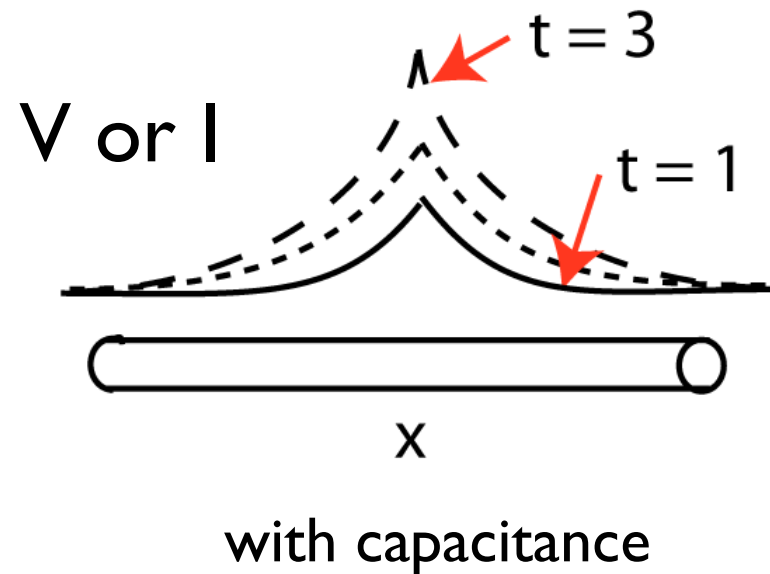
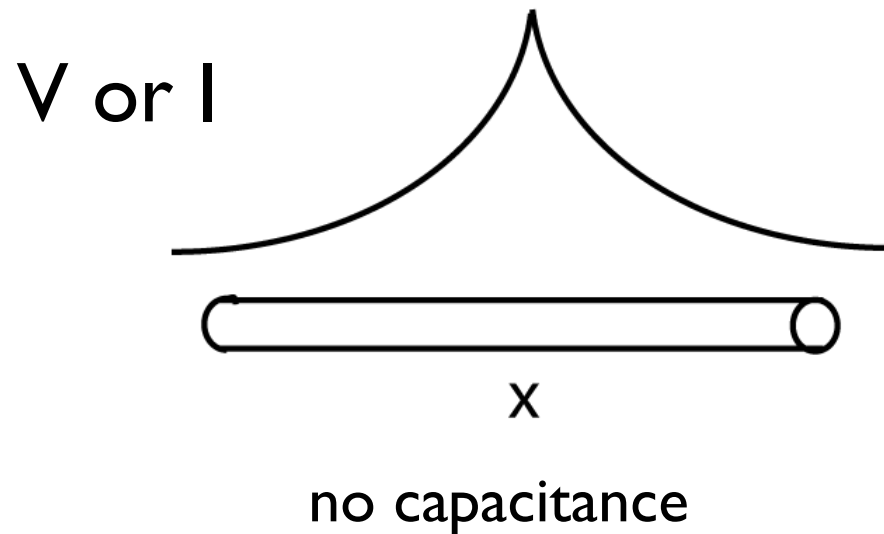
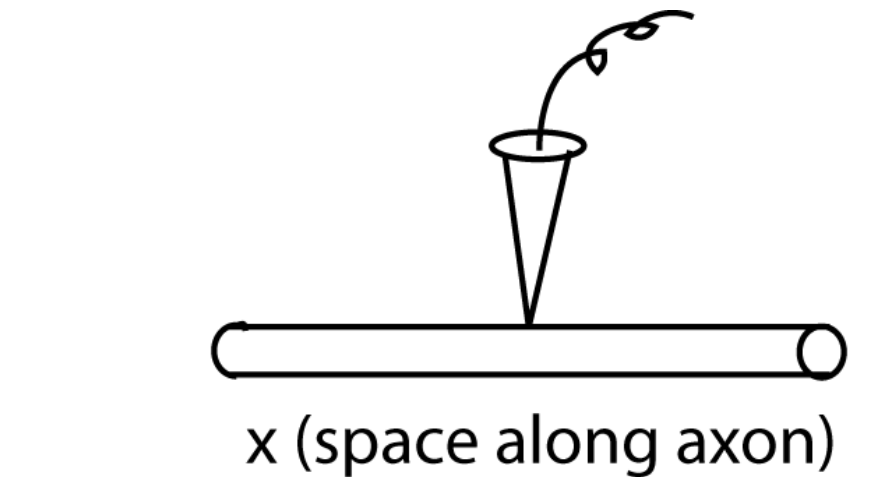
patch of membrane with  
Nernst potential across  
channel - battery

# What happens if we inject current into dendrite?



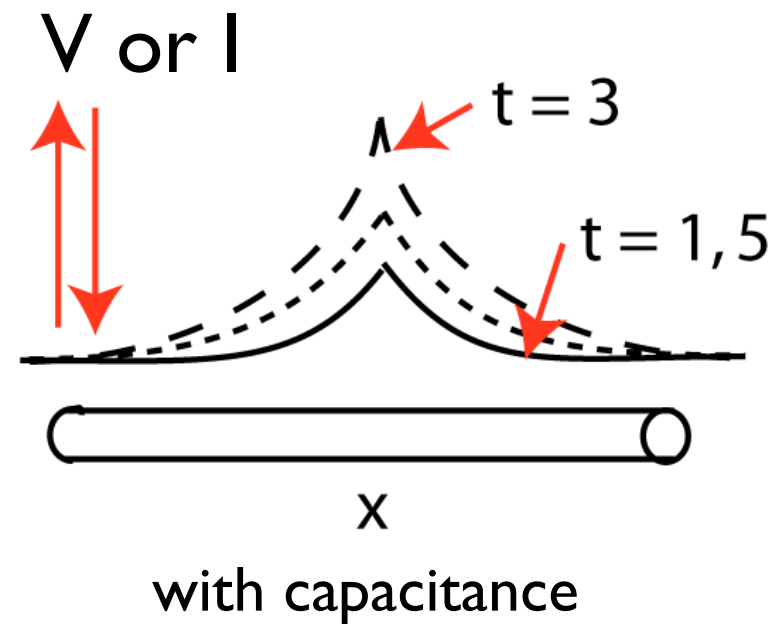
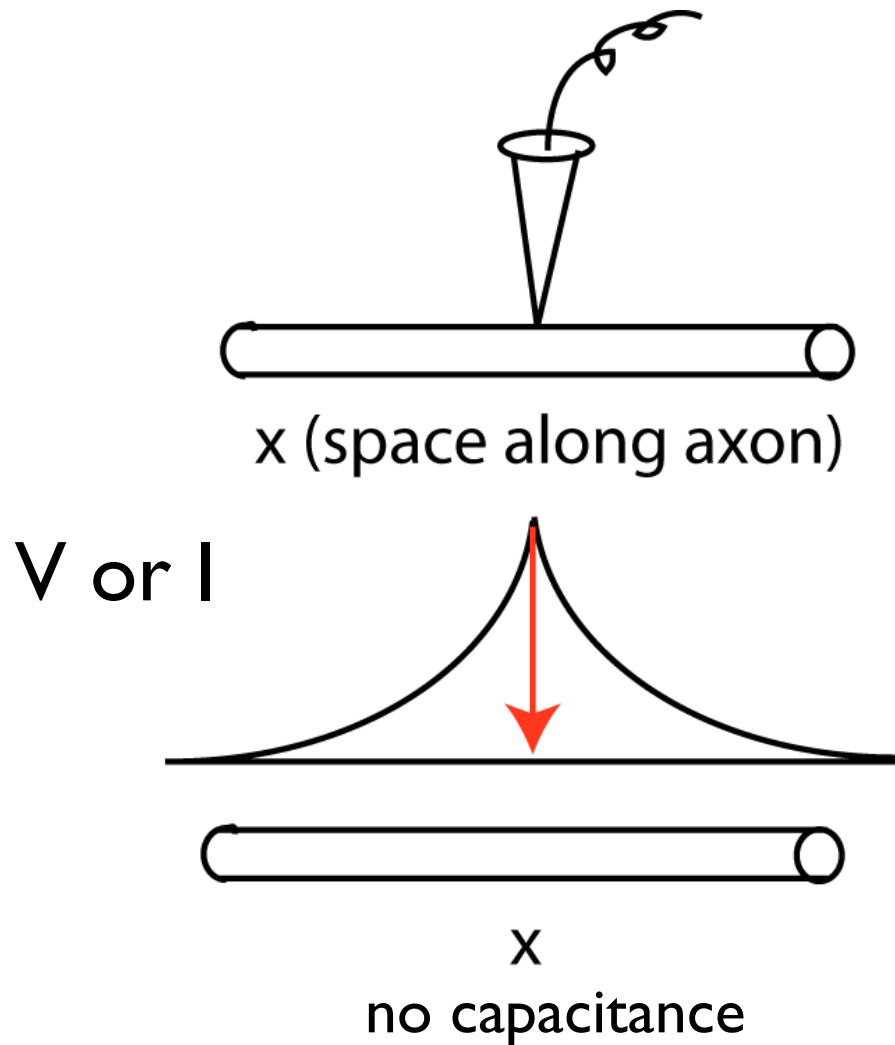
Current will start to flow everywhere, following the path of least resistance

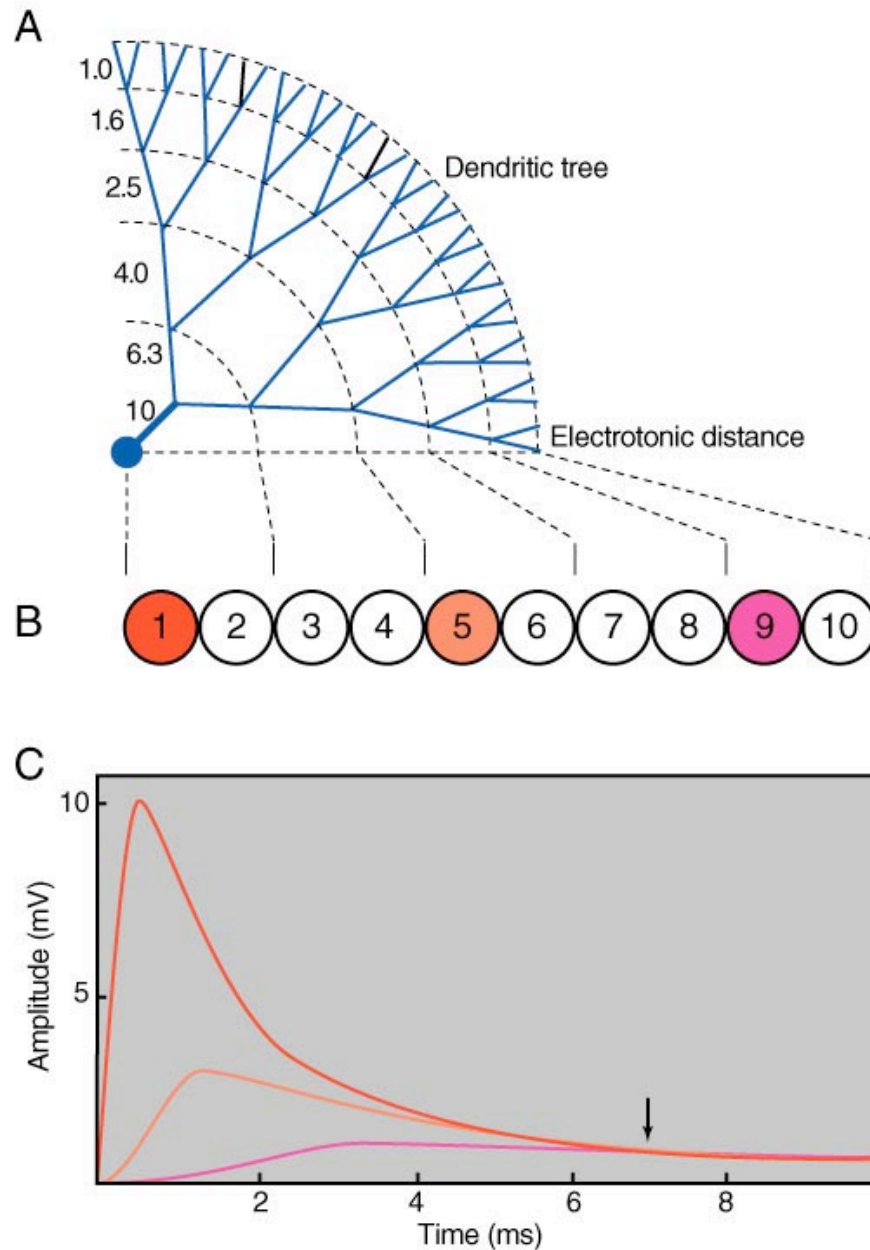
# Steady-state current: with and without capacitance





# Transient impulse: with and without capacitance

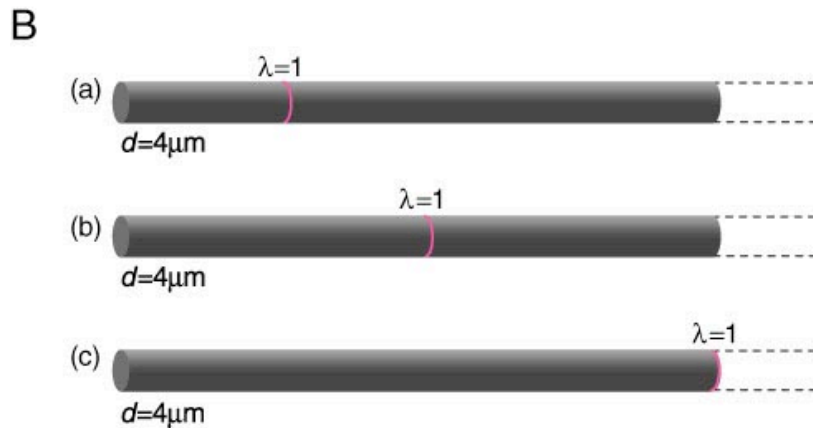
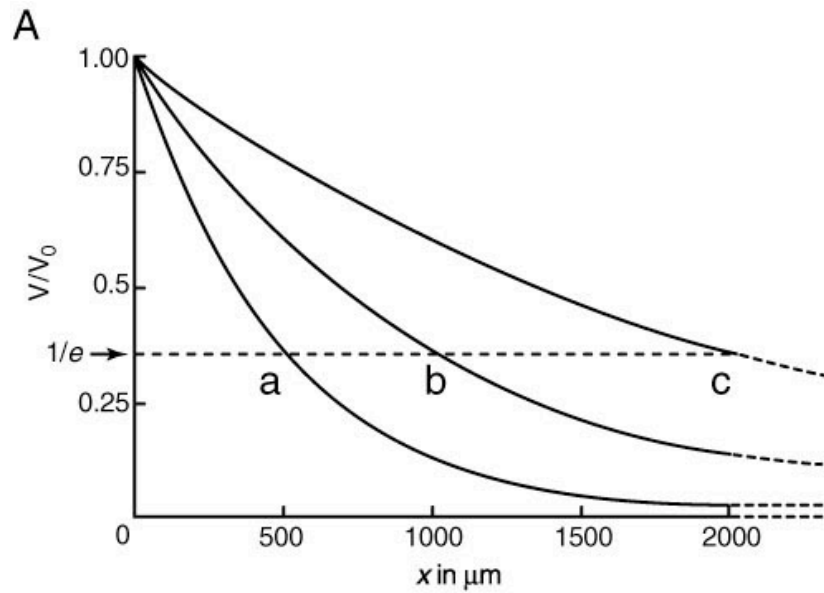




- spread of electrotonic potentials is delayed and of smaller amplitude the farther away from injection site

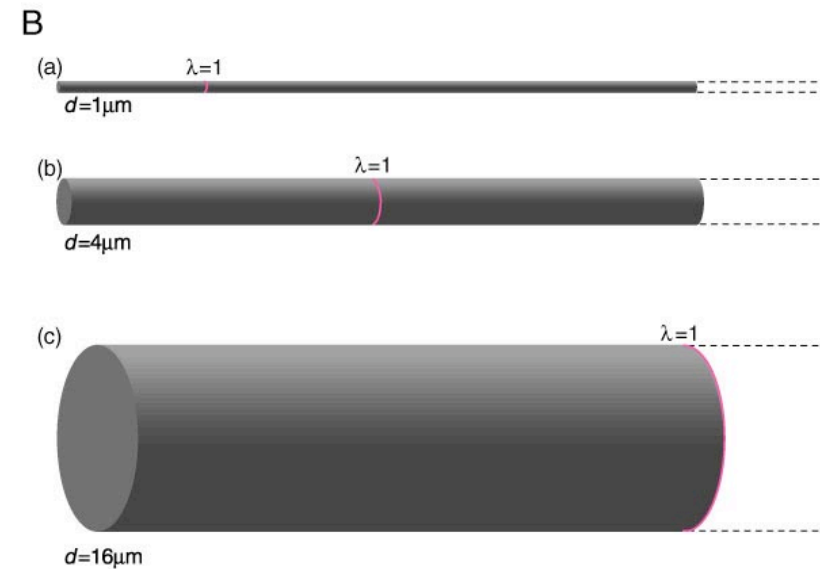
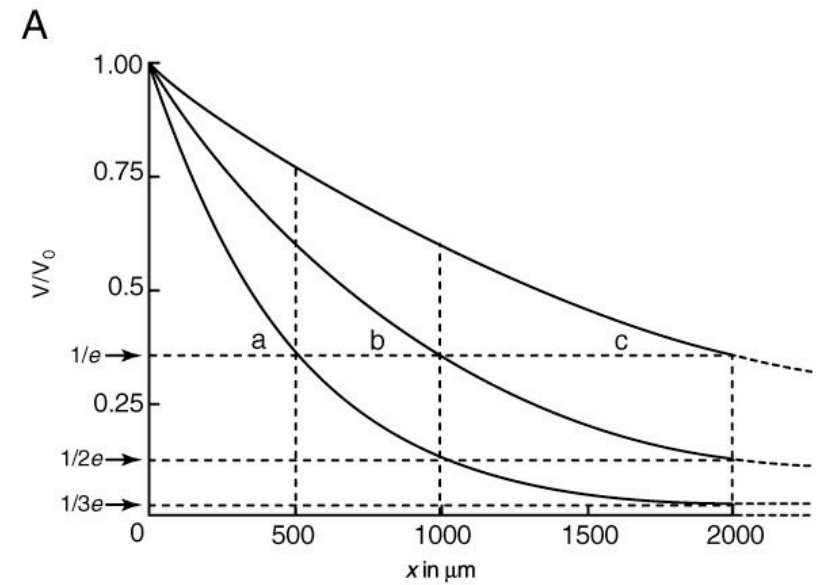
# Length constant

- characteristic length (membrane space constant)  $\lambda$  (lambda) - depends on  $R_m$  and  $R_L$  (also on diameter of process - big diameter, low  $R_L$ )
- the length of dendrite over which the electrotonic potential decays to a value of 0.37 of value at injection site



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high  $R_m$  and low  $R_L$   
increase  $\lambda$



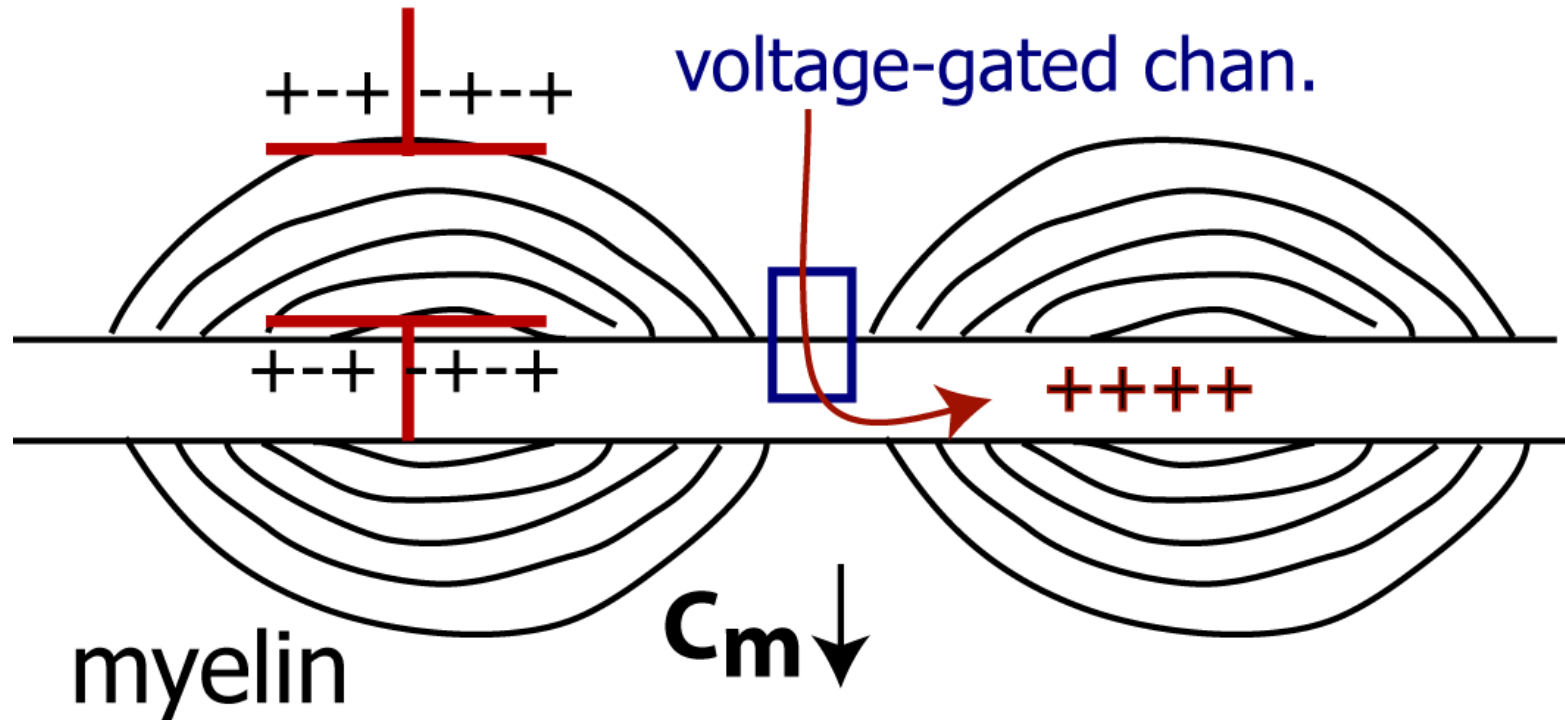
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big diameter  
increases  $\lambda$

# Time constant $\tau$

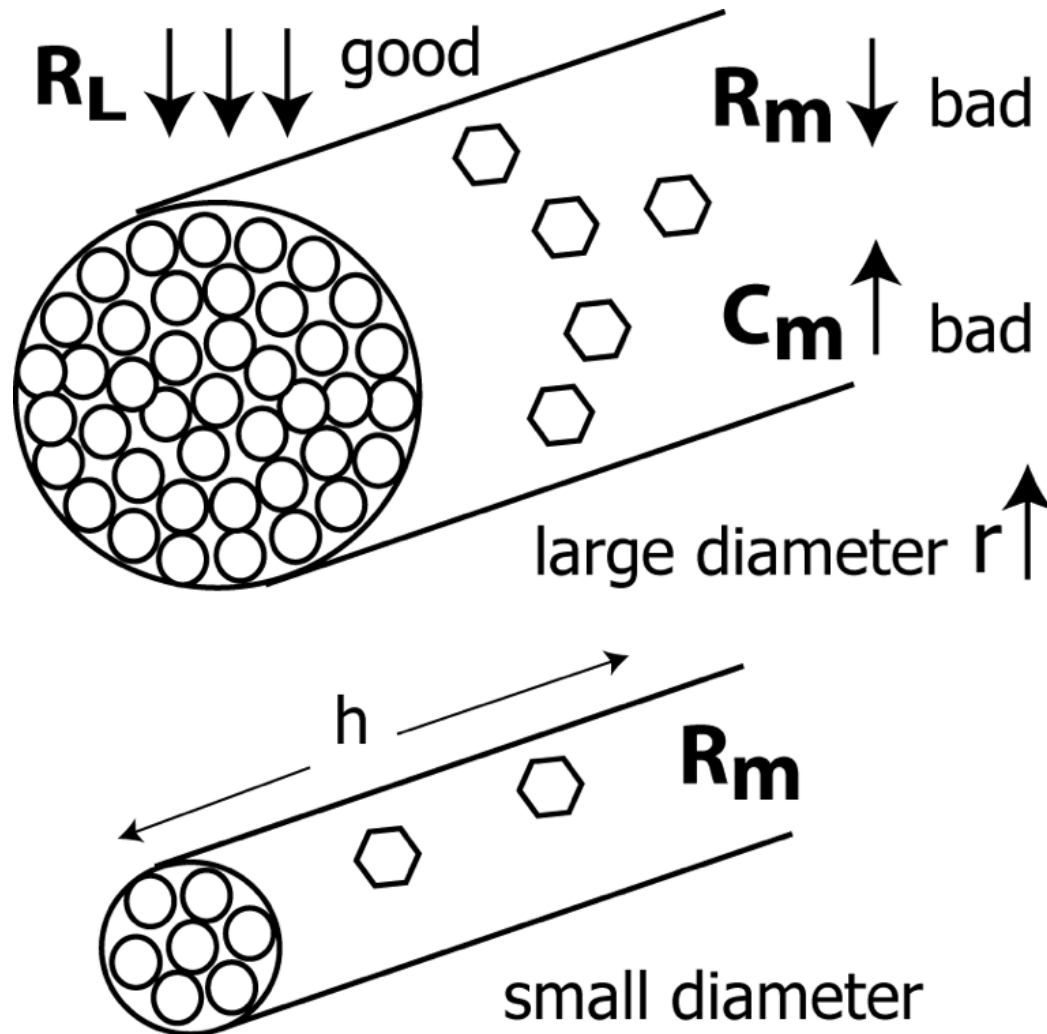
- membrane time constant  $\tau$  (tau) depends on  $C_m$
- the time required for voltage change across membrane to reach 0.37 of its final value (i.e. of maximally charged capacitor)
- the greater the capacitance, the greater  $\tau$  is

# Myelin decreases capacitance



- \* Myelin separates the plates of the capacitor - current won't get wasted charging up the capacitor
- \* (myelin also INcreases  $R_m$  - less leakage)

# Increasing diameter of axon



$$\text{volume} = \pi r^2 * h$$

$$\text{surf. area} = 2 \pi r * h$$

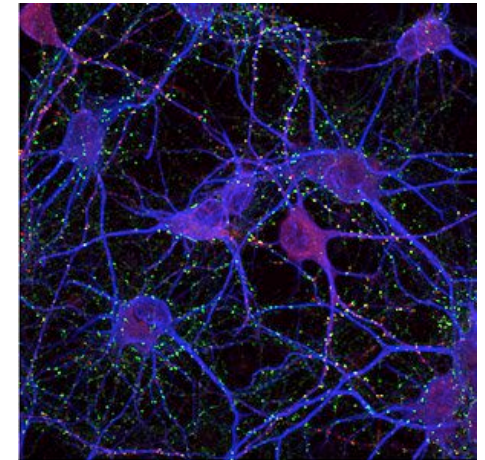
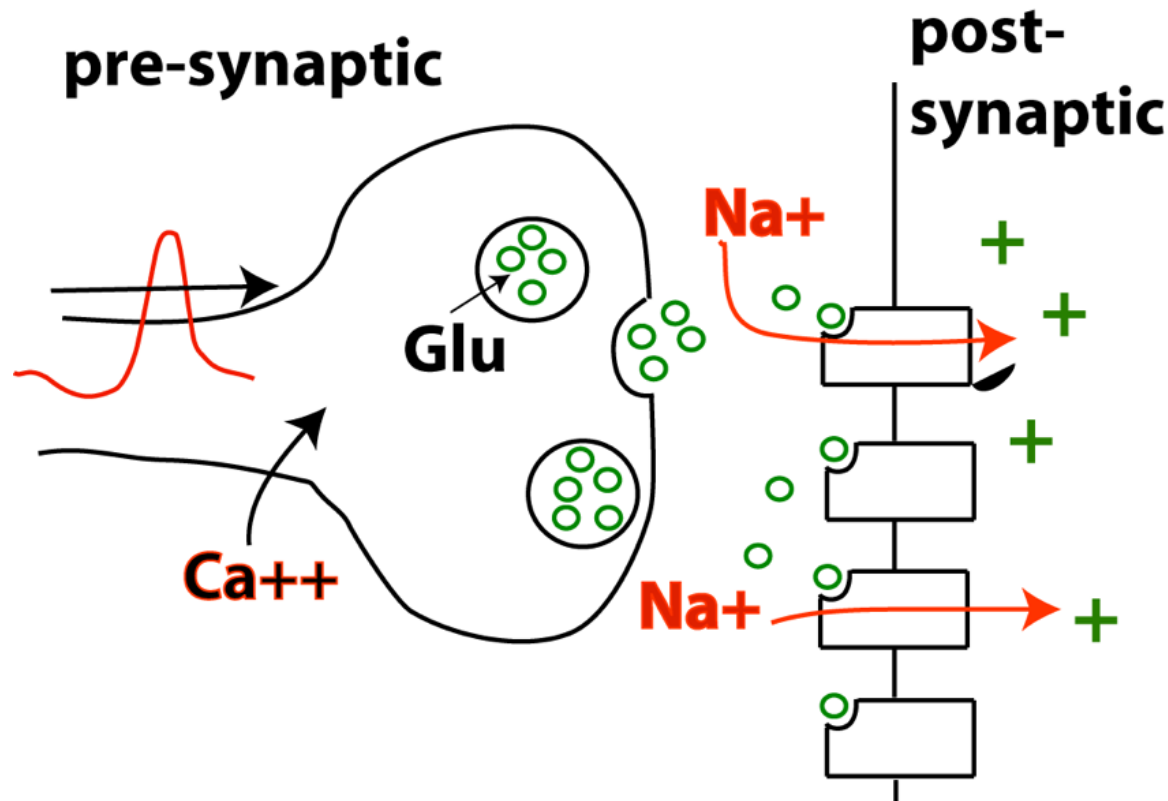
→ volume goes up faster than membrane surface area with increased diameter

→ decrease in longitudinal resistance greater than increase in  $C_m$  or decrease in  $R_m$

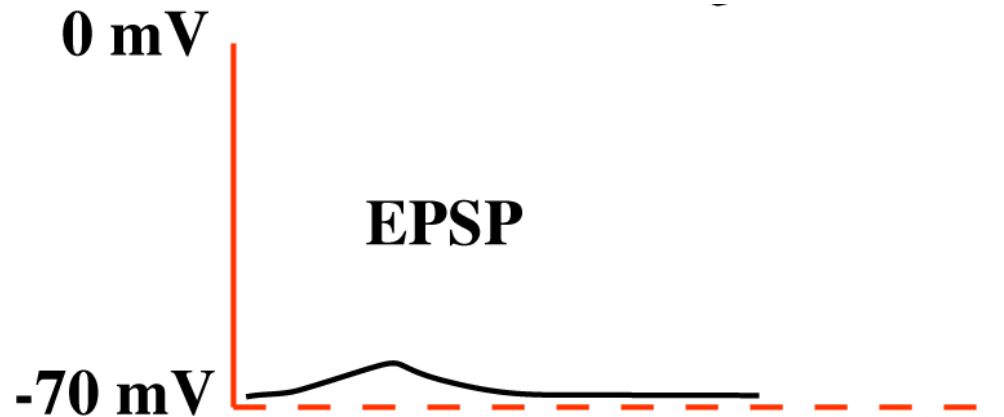


- in order to spread electrotonic potentials as far as possible, we want:
  - high membrane resistance (myelin)
  - low membrane capacitance (myelin)
  - low internal resistance (large diameter)

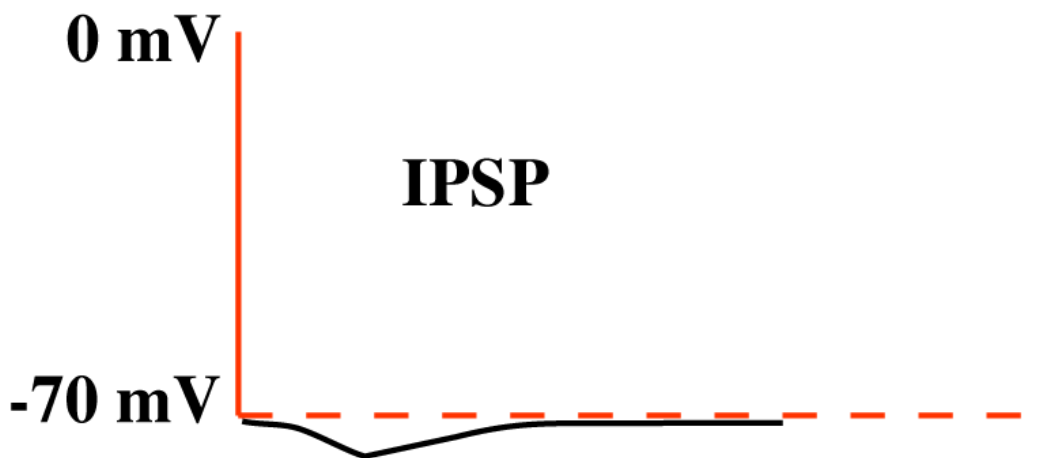
# Synaptic Transmission



# EPSP and IPSP



- excitatory post-synaptic pot. (EPSP)

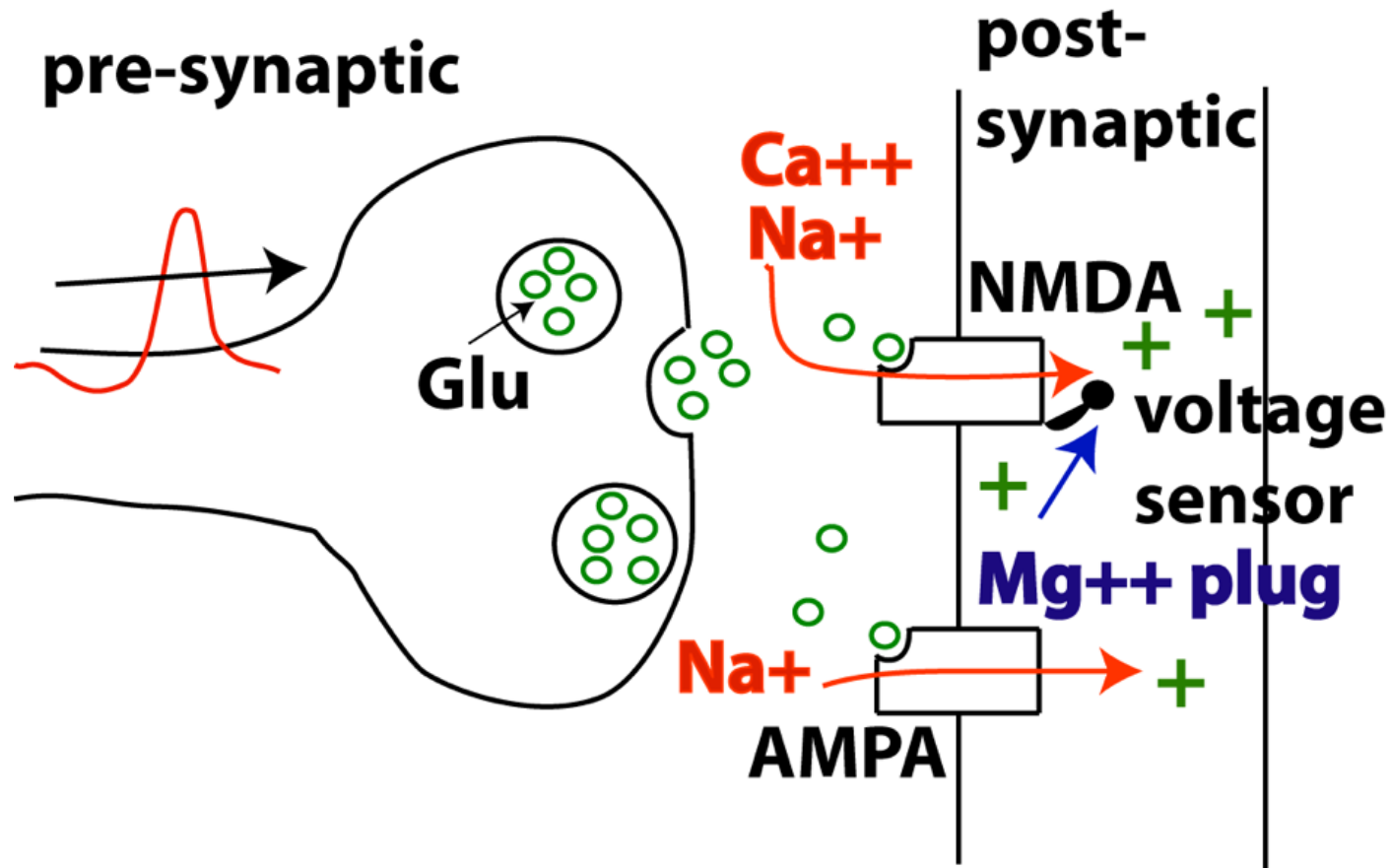


- inhibitory post-synaptic pot. (IPSP)
- one input not enough

# Receptor channels (examples)

- **AMPA** (alpha-amino-3-hydroxy-5-methylisoxazole-4-propionic acid) - **excitatory**;  $E_{\text{AMPA}} = \sim -10 \text{ mV}$ ; NT = Glu; conducts  $\text{Na}^+$ ,  $\text{Ca}^{++}$
- **NMDA** (N-methyl-D-aspartic acid) - **excitatory**; NT = Glu, voltage-sensitive;  $E_{\text{NMDA}} = 0 \text{ mV}$ ; conducts  $\text{Na}^+$ ,  $\text{Ca}^{++}$ ,  $\text{K}^+$
- **GABA<sub>A</sub>** (Gamma-aminobutyric acid): - **inhibitory**; NT = GABA;  $E_{\text{GABA}_A} = -70 \text{ mV}$ ; conducts  $\text{Cl}^-$
- **GABA<sub>B</sub>**: - **inhibitory**; NT = GABA,  $E_{\text{GABA}_B} = \sim -80 \text{ mV}$ ; conducts  $\text{K}^+$

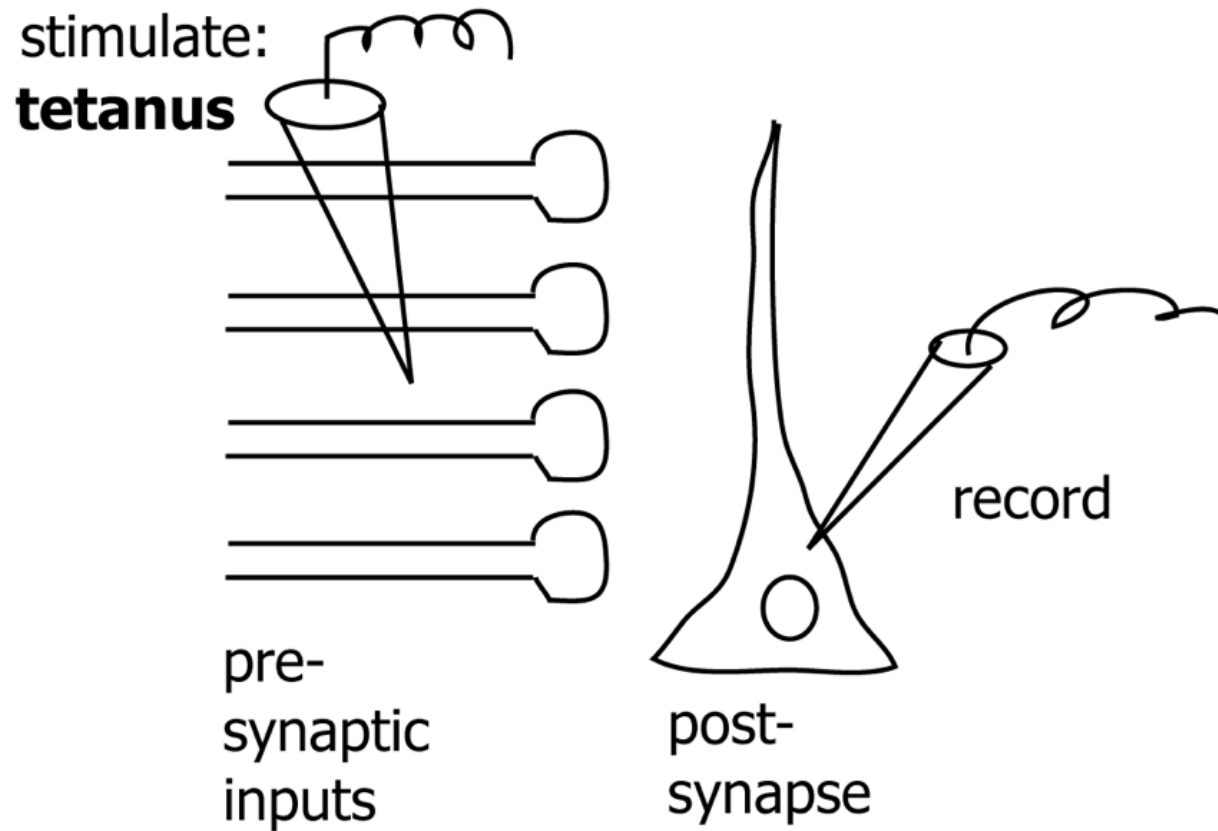
# NMDA channels act as AND gates



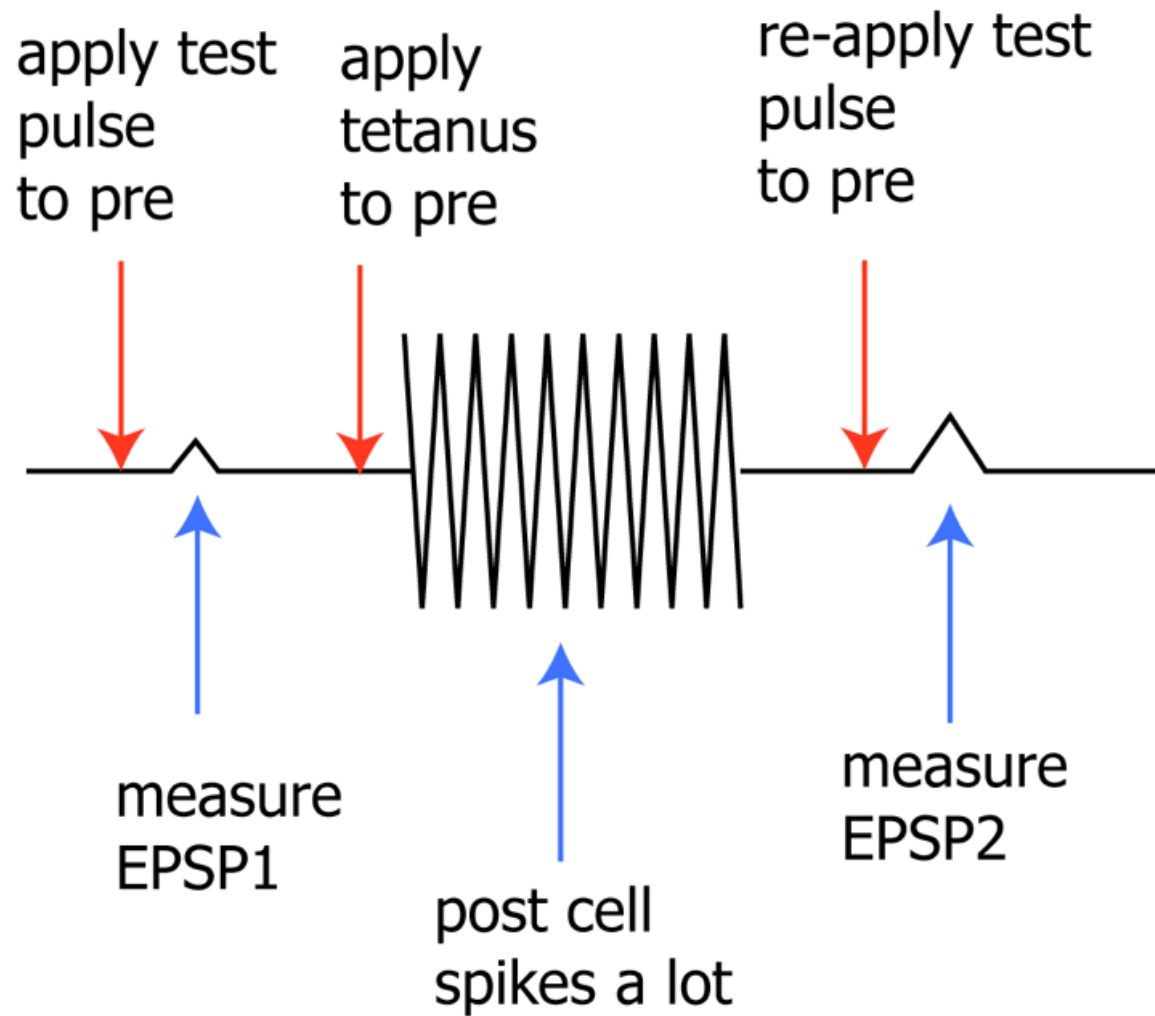
NMDA requires both depolarization AND glutamate

# LTP

- long term potentiation

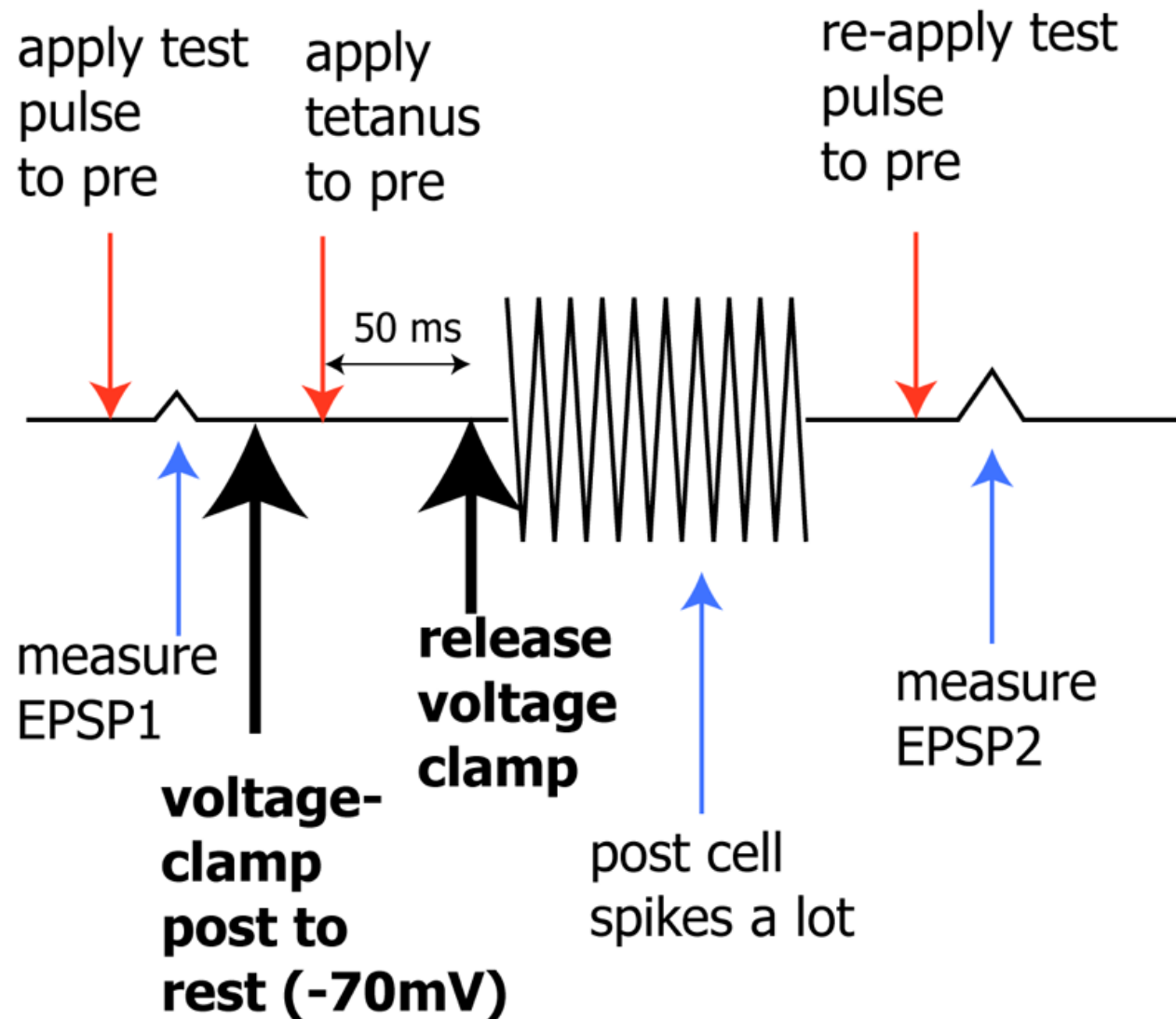


# Inducing and measuring LTP



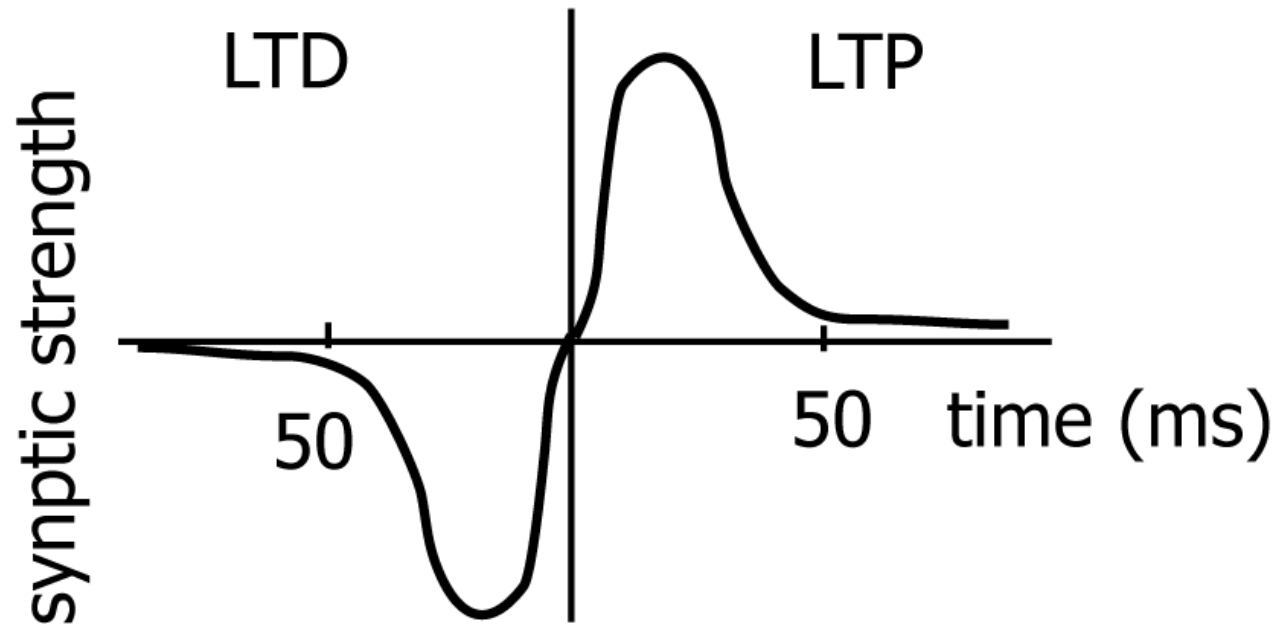
If  $EPSP2 > EPSP1$ , LTP has occurred

# Timing of pre-synaptic stimulation and post-synaptic response matters





# Spike-timing dependent plasticity (STDP)



# Synaptic strength change

- if pre spikes within 50 ms before post: LTP
- if post spikes within 50 ms before pre: LTD
- if pre and post spike  $> 50$  ms apart: no change

# Possible LTP mechanisms

