



## Minireview

## Role of adiponectin system in insulin resistance



Chiara Caselli\*

Consiglio Nazionale delle Ricerche (CNR), Institute of Clinical Physiology, Laboratory of Cardiovascular Biochemistry, Pisa 56100, Italy

## ARTICLE INFO

## Article history:

Received 15 July 2014

Received in revised form 4 September 2014

Accepted 4 September 2014

Available online 16 September 2014

## Keywords:

Adiponectin

Adiponectin signaling

Insulin resistance

Therapeutic target

## ABSTRACT

The knowledge of the pathogenesis of obesity and its metabolic sequelae has significantly advanced over the last few decades and adipose tissue is now considered a link between obesity and insulin resistance. Adiponectin, one of the major adipocyte-secreted proteins, has attracted scientific interest in recent years and has been extensively studied both in human and animal models. Adiponectin exerts insulin-sensitizing effects through binding to its receptors, leading to activation of AMPK, PPAR- $\alpha$ , and potentially other unknown molecular pathways. In obesity-linked insulin resistance, both adiponectin and adiponectin receptors are downregulated, leading to activation of signaling pathways involved in metabolism regulation. Up-regulation of adiponectin/adiponectin receptors or enhancing adiponectin receptor function may be an interesting therapeutic strategy for obesity-linked insulin resistance. In this review we will focus on the recent research related to the relationship between the adiponectin system and insulin resistance. The potential use of adiponectin or its receptor for therapeutic intervention will be also discussed.

© 2014 Elsevier Inc. All rights reserved.

## Contents

1. Obesity and insulin resistance . . . . .	155
2. Adiponectin . . . . .	156
2.1. Structure and biosynthesis . . . . .	156
2.2. Gene transcription . . . . .	156
2.3. Adiponectin receptors . . . . .	156
2.4. Adiponectin and insulin resistance . . . . .	156
2.5. Adiponectin as therapeutic target . . . . .	158
3. Conclusion . . . . .	159
4. Conflict of interest . . . . .	159
References . . . . .	159

## 1. Obesity and insulin resistance

In Western countries, obesity is reaching epidemic proportions and is associated with a high prevalence of type 2 diabetes mellitus (T2DM) characterized by insulin resistance of peripheral tissues such as liver, muscle, and fat [1]. In 80% of cases the presence of T2DM was related to obesity [2]. Our understanding of the pathogenesis of obesity and its metabolic sequelae has significantly advanced over the past few decades, and adipose tissue is now considered a link between obesity and insulin resistance [3,4].

Adipose tissue is now regarded as not just a storage depot for excess energy, but rather as an endocrine organ, secreting a large number of bioactive molecules called adipokines [5]. This family of cytokines includes molecules with high biological activity, such as adiponectin, resistin, leptin, and PAI-1 (plasminogen activator inhibitor-1). These adipose-tissue-derived factors show paracrine activity, sustaining the inflammatory condition of adipose tissue, as well as endocrine activity, having effects on metabolism and inflammation [6–8].

Adiponectin is an abundantly expressed adipokine that exerts a potent insulin-sensitizing effect through binding to its receptors. In obesity-linked insulin resistance, both adiponectin and adiponectin receptors are downregulated, leading to activation of signaling pathways involved in metabolism regulation [9–12]. Upregulation of

\* CNR, Institute of Clinical Physiology, Via Moruzzi 1, 56124 Pisa, Italy.  
E-mail address: [chiara.caselli@ifc.cnr.it](mailto:chiara.caselli@ifc.cnr.it).

adiponectin/adiponectin receptors or enhancing adiponectin receptor function may be an interesting therapeutic strategy for obesity-linked insulin resistance.

## 2. Adiponectin

Adiponectin is a major adipocyte-secreted protein and is down-regulated in obesity and its related pathology [9,11]. In contrast to the other adipokines, adiponectin exerts antidiabetic, anti-atherogenic and anti-inflammatory activities [12–18]. Due to these positive actions, adiponectin has attracted tremendous scientific interest in recent years, and has been extensively studied both in human and animal models. The structure, biosynthesis, and signaling of adiponectin in muscle and liver are illustrated in Fig. 1.

### 2.1. Structure and biosynthesis

The gene encoding human adiponectin, also called Acrp30, AdipoQ, apM1 or GBP28, is located on chromosome 3q27 [19], a locus linked with susceptibility to diabetes and cardiovascular disease [20]. The full-length protein consists of 247 amino acids, including the N-terminal hypervariable region, a conserved collagenous domain comprising 22 Gly-Xaa-Yaa repeats and a C-terminal C1q-like globular domain [21]. Adiponectin is present in peripheral circulation as three oligomeric complexes. The trimeric adiponectin represents the basic unit and is called low-molecular-weight (LMW) adiponectin. Two subunits of the trimer are linked by the collagen-like domain to form a hexamer, also termed middle-molecular-weight (MMW) adiponectin. Hexamer subunits are linked in a bouquet-like high-molecular-weight (HMW) adiponectin [22–24]. Moreover, cleavage of the full-length form generates the 17 kDa globular fragment of adiponectin, called globular adiponectin, which is found at lower levels (about 1% of total adiponectin) in the circulation [25].

Multimeric complex formation of adiponectin is recognized as an important mechanism modulating its biological functions [24]. Both *in vitro* and *in vivo* studies have suggested that HMW is the biologically active form and that HMW, rather than total adiponectin, may exert anti-atherogenic, anti-diabetic and anti-inflammatory actions that could prevent the development of metabolic and cardiovascular disease [26].

The biosynthesis and secretion of adiponectin in adipocytes are regulated in the endoplasmic reticulum [27–29]. Post-translational modifications are required for intracellular assembly of the HMW oligomeric complex in adipocytes, for its secretion, and also for maintaining its stability in the circulation [30]. The development of ELISA methods contributed to a more widespread measure of the adiponectin levels in peripheral circulation, thus increasing the experimental data needed to confirm its role in different patho-physiological conditions, especially in cardiovascular disease [31].

### 2.2. Gene transcription

The adiponectin gene promoter contains multiple transcription factor binding sites through which a large number of diverse factors have been shown to modulate its activity [32,33]. Adiponectin gene expression is upregulated by transcription factors such as peroxisome proliferator activator receptor- $\gamma$  (PPAR- $\gamma$ ) [34], C/EBP $\alpha$ , and Forkhead transcription factor FoxO1 [35,36]. Adiponectin gene expression is downregulated in the adverse environment associated with obesity, such as chronic low-grade inflammation and oxidative stress [33]. It is regulated through the Akt and JAK/STAT signaling pathways [37] by oxidative stress, through protein kinase C [38] and through the JNK signaling pathway by TNF $\alpha$  [39], and through p44/42 MAPK by IL-6 [40]. Moreover, under conditions of endoplasmic reticulum stress, adiponectin mRNA expression is downregulated [41,42]. Other transcription factors known to downregulate adiponectin include CREB,

which activates ATF3, NFAT, and protein kinase A, which is stimulated by beta-adrenergic signaling [32,33].

### 2.3. Adiponectin receptors

The two main adiponectin receptors, AdipoR1 and AdipoR2, are structurally and functionally distinct from classic G-protein-coupled receptors. They contain seven transmembrane domains, have an inverted membrane topology with a cytoplasmic N-terminus and a short extracellular C-terminus of approximately 25 amino acids [43,44]. AdipoR1 is expressed ubiquitously, whereas AdipoR2 is expressed most abundantly in the liver [44].

Recently, molecules that couple the AdipoRs to their downstream signaling cascades have been identified, including the adaptor protein containing a PH (pleckstrin homology) domain (APPL1), receptor for activated protein kinase C1 (RACK1) [45–47], protein kinase CK2 $\beta$ , and endoplasmic reticulum protein of 46 kDa (ERp46) [48–51]. However, detailed molecular events remain elusive.

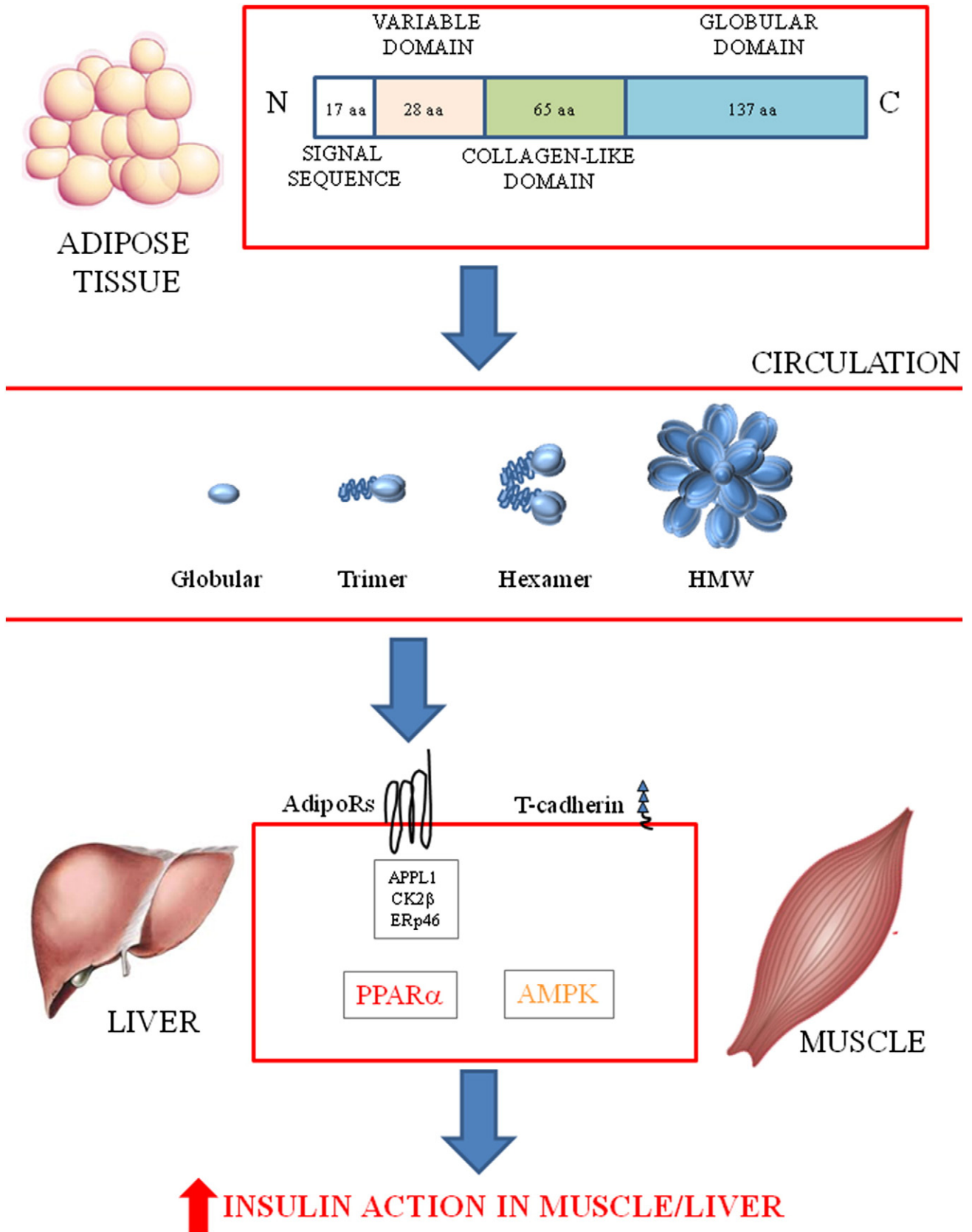
In addition to AdipoRs, T-cadherin has also been suggested as a potential receptor for adiponectin [52]. T-cadherin is different from classical cadherin molecules due to the lack of a cytoplasmic domain (so T-cadherin is designated as truncated cadherin), and the presence of a highly conserved amino acid motif throughout evolution in vertebrates [53,54]. T-cadherin is highly expressed in the heart, smooth muscle and endothelium, representing the main targets of adiponectin. It is abundantly expressed in injured vascular endothelial and smooth muscle cells in atherosclerotic regions [55,56] and it has been shown to protect against cardiac stress [57]. However, the molecular mechanisms of transmission of the adiponectin signal and the functional relevance of this binding remain unclear and require more detailed studies.

Finally, similar to other members of the collectin family, adiponectin binds to calreticulin on the cell surface of macrophages, facilitating the uptake of dead cells [58,59]. These data suggest that adiponectin can protect the organism from systemic inflammation at least in part through its ability to function as a collectin protein [59].

### 2.4. Adiponectin and insulin resistance

AdipoRs are mainly involved in AMP-activated protein kinase (AMPK) and peroxisome-proliferator-activated receptor  $\alpha$  (PPAR- $\alpha$ ) ligand activities [44]. Adiponectin exerts effects on glucose uptake and  $\beta$ -oxidation via AMPK [60]. In skeletal muscle, activation of AMPK is stimulated by globular and full-length adiponectin, while in the liver only by the full-length form [60,61]. The glucose-lowering effects of adiponectin may account for the phosphorylation of acetyl coenzyme-A carboxylase (ACC), increased fatty-acid combustion, glucose uptake and lactate production in myocytes. These activities stimulated by adiponectin limit gluconeogenesis in the liver. Via PPAR- $\alpha$ , adiponectin increases fatty-acid and energy consumption, leading to reduced triglyceride content and increased insulin sensitivity in the liver and skeletal muscle [62].

Studies in animal models have confirmed the importance of AdipoRs in mediating physiologically metabolic regulation by adiponectin, and have highlighted their functional differences [63–65], also reporting conflicting data. The adiponectin-induced activation of AMPK is blocked in mice by an AdipoR1-targeted deletion; disruption of AdipoR2 decreased adiponectin-stimulated PPAR- $\alpha$  signaling; and simultaneous ablation of both AdipoR1 and AdipoR2 blocked the binding and actions of adiponectin, leading to insulin resistance and glucose intolerance [63]. AdipoR1-null mice showed increased adiposity and glucose intolerance, whereas AdipoR2-null mice were lean and resistant to diet-induced glucose intolerance, indicating that AdipoR1 and AdipoR2 might have opposing effects [64]. By contrast, deletion of AdipoR2 reduced diet-induced insulin resistance, but increased their susceptibility to T2DM [65].



**Fig. 1.** Adiponectin biosynthesis from adipose tissue, its release into bloodstream, and its mechanisms of action on target tissues, such as muscle and liver. Adiponectin molecules are produced by adipocytes and, after post-translational modifications, globular, trimer, hexamer and HMW isoforms of adiponectin are secreted in peripheral circulation. After their binding to AdipoRs on target tissues, adiponectin exerts effects on glucose uptake and  $\beta$ -oxidation via AMPK and it increases fatty-acid and energy via PPAR- $\alpha$ . Collectively, these actions lead to improving insulin action in liver and muscle.

Besides the insulin-sensitizing effect of adiponectin by linking to AMPK and PPAR- $\alpha$ , adiponectin decreases tissue triglyceride levels and increases insulin signaling. In skeletal muscle, adiponectin increases fatty-acid transport, stimulating the expression of proteins such as CD36, acyl-coenzyme A oxidase [66]. On the contrary, high tissue triglyceride levels could interfere with glucose uptake through activation of insulin-stimulated phosphatidylinositol (PI) 3-kinase as well as translocation of glucose transporter-4 (GLUT-4), thus leading to the development of insulin resistance.

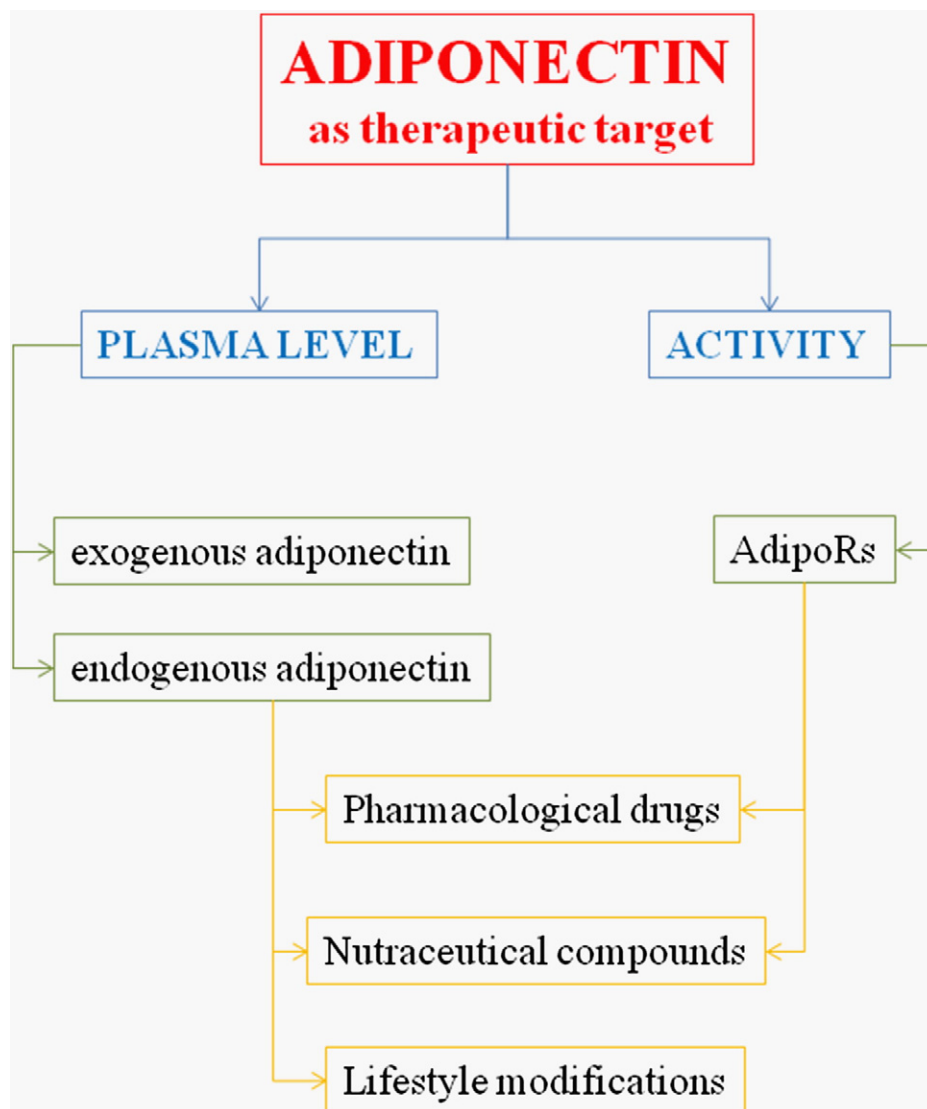
### 2.5. Adiponectin as therapeutic target

Several strategies have been suggested for increasing the favorable effects of adiponectin, including enhancing both its plasma level and activities (Fig. 2).

Adiponectin circulating levels could be elevated either by directly using exogenous adiponectin, e.g., through an injection, or by increasing the endogenous adiponectin through treatments. Because of the high circulating levels and multimeric conformations of adiponectin, the direct use of exogenous adiponectin is difficult. Thus, the best option remains increasing endogenous adiponectin through the use of pharmacological

drugs, nutraceutical compounds and lifestyle modification. Pharmaceutical products effective in elevating adiponectin circulating levels include the PPAR- $\alpha$  agonists thiazolidinediones (TZDs) [67–71], inhibitors of the renin-angiotensin system such as angiotensin converting enzyme (ACE) inhibitors and angiotensin II receptor blockers (ARBs) [72,73], and statins, although their role is still debated. Among nutraceutical compounds, fish oil, linoleic acid, seed extract, green tea extract, and polyphenol resveratrol are able to elevate adiponectin concentration [74]. Moreover, weight loss or physical activity is able to increase adiponectin levels, especially among obese or diabetic subjects [75].

An alternative approach for increasing favorable adiponectin effects is enhancing its signaling through compounds that can act on AdipoR. Agonists of PPAR- $\alpha$ , PPAR- $\gamma$  and ARBs are able to up-regulate expression of AdipoR [76]. Moreover, a member of the plant defense proteins, osmotin, has been recently identified as a potential adiponectin receptor agonist [77]. Very recently, an orally active synthetic small-molecule AdipoR agonist (AdipoRon) was identified. In animal models, AdipoRon was able to ameliorate obesity-related diseases such as insulin resistance, glucose intolerance, and T2DM, and to prolong lifespan [78]. Due to the presence of AdipoR1 dimers in various cell lines as well as the regulation of this dimerization by adiponectin, the



**Fig. 2.** Summary of the potential strategies that could be used to increase adiponectin effects. The positive effects of adiponectin could be obtained either rising directly the adiponectin circulating levels (supplementing with exogenous adiponectin as well as by increasing the release of endogenous adiponectin), or increasing indirectly its activity at target tissue level. Pharmacological drugs, nutraceutical compounds, and lifestyle modification could represent reliable options able to therapeutically target adiponectin system.

oligomerization of AdipoRs could be considered a potential pharmacological target in obesity-associated disorders [79,80].

### 3. Conclusion

In recent years, due to its positive regulatory action in several conditions including insulin resistance, the adiponectin system has become critical for designing new drugs. Several issues linked to the molecular and cellular mechanisms underlying the adiponectin system could be taken into account as potentially useful targets in new pharmacological approaches. These include the biosynthetic pathway of adiponectin; the multimerization process, in order to increase the HMW adiponectin, considered the biologically active form; the modulation of adiponectin levels and/or activity at gene level; adiponectin signaling, including both AdipoRs and downstream post-receptor mechanisms; and the protective activities of adiponectin at the intracellular level, in order to develop adiponectin-based therapeutics with cell-specific delivery approaches.

In conclusion, further work is needed to completely elucidate the molecular mechanisms of biosynthesis, secretion and signaling of adiponectin and their potential therapeutic value. Moreover, biologically active adiponectin forms or chemical entities that can activate adiponectin receptors should be additionally investigated and applied in human trials to test their potential benefit in clinical settings.

### 4. Conflict of interest

The author declares that she has no conflict of interest.

### References

- [1] A.R. Saltiel, The molecular and physiological basis of insulin resistance: emerging implications for metabolic and cardiovascular diseases, *J. Clin. Invest.* 106 (2000) 163–45.
- [2] S. Matthaehi, M. Stumvoll, M. Kellerer, H.U. Häring, Pathophysiology and pharmacological treatment of insulin resistance, *Endocr. Rev.* 21 (2000) 585–618.
- [3] Y. Li, L. Ding, W. Hassan, D. Abdelkader, J. Shang, Adipokines and hepatic insulin resistance, *J. Diabetes Res.* 2013 (2013) 170532.
- [4] K. Rabe, M. Lehrke, K.G. Parhofer, U.C. Broedl, Adipokines and insulin resistance, *Mol. Med.* 14 (2008) 741–751.
- [5] H. Tilg, A.R. Moschen, Adipocytokines: mediators linking adipose tissue, inflammation and immunity, *Nat. Rev. Immunol.* 6 (2006) 772–783.
- [6] Y. Matsuzawa, The metabolic syndrome and adipocytokines, *FEBS Lett.* 580 (2006) 2917–2921.
- [7] F. Lago, C. Dieguez, J. Gómez-Reino, O. Gualillo, Adipokines as emerging mediators of immune response and inflammation, *Nat. Clin. Pract. Rheumatol.* 3 (2007) 716–724.
- [8] D.C. Lau, B. Dhillon, H. Yan, P.E. Szmitko, S. Verma, Adipokines: molecular links between obesity and atherosclerosis, *Am. J. Physiol. Heart Circ. Physiol.* 288 (2005) H2031–H2041.
- [9] Y. Arita, S. Kihara, N. Ouchi, M. Takahashi, K. Maeda, J. Miyagawa, et al., Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity, *Biochem. Biophys. Res. Commun.* 425 (2012) 560–564.
- [10] T. Kadowaki, T. Yamauchi, N. Kubota, K. Hara, K. Ueki, K. Tobe, Adiponectin and adiponectin receptors in insulin resistance, diabetes, and the metabolic syndrome, *J. Clin. Invest.* 116 (2006) 1784–1792.
- [11] A.S. Lihn, S.B. Pedersen, B. Richelsen, Adiponectin: action, regulation and association to insulin sensitivity, *Obes. Rev.* 6 (2005) 13–21.
- [12] A. Yadav, M.A. Kataria, V. Saini, A. Yadav, Role of leptin and adiponectin in insulin resistance, *Clin. Chim. Acta* 417 (2013) 80–84.
- [13] B.J. Goldstein, R.G. Scalia, X.L. Ma, Protective vascular and myocardial effects of adiponectin, *Nat. Clin. Pract. Cardiovasc. Med.* 6 (2009) 27–35.
- [14] A.H. Berg, T.P. Combs, X. Du, M. Brownlee, P.E. Scherer, The adipocyte-secreted protein Acrp30 enhances hepatic insulin action, *Nat. Med.* 7 (2001) 947–953.
- [15] T. Yamauchi, J. Kamon, H. Waki, Y. Terauchi, N. Kubota, K. Hara, et al., The fat-derived hormone adiponectin reverses insulin resistance associated with both lipotrophy and obesity, *Nat. Med.* 7 (2001) 941–946.
- [16] T. Yokota, K. Oritani, I. Takahashi, J. Ishikawa, A. Matsuyama, N. Ouchi, et al., Adiponectin, a new member of the family of soluble defense collagens, negatively regulates the growth of myelomonocytic progenitors and the functions of macrophages, *Blood* 96 (2000) 1723–1732.
- [17] N. Ouchi, K. Walsh, Adiponectin as anti-inflammatory factor, *Clin. Chem. Acta* 380 (2007) 24–30.
- [18] N. Ouchi, R. Shibata, K. Walsh, Cardioprotection by adiponectin, *Trends Cardiovasc. Med.* 16 (2006) 141–146.
- [19] K. Saito, T. Tobe, S. Minoshima, S. Asakawa, J. Sumiya, M. Yoda, et al., Organization of the gene for gelatin-binding protein [GBP28], *Gene* 229 (1999) 67–73.
- [20] M. Stumvoll, O. Tschritter, A. Fritsche, H. Staiger, W. Renn, M. Weisser, et al., Association of the T-G polymorphism in adiponectin [exon 2] with obesity and insulin sensitivity: interaction with family history of type 2 diabetes, *Diabetes* 51 (2002) 37–41.
- [21] Y. Wang, K.S. Lam, M.H. Yau, A. Xu, Post-translational modifications of adiponectin: mechanisms and functional implications, *Biochem. J.* 409 (2008) 623–633.
- [22] H. Ebinuma, T. Miida, T. Yamauchi, Y. Hada, K. Hara, N. Kubota, et al., Improved ELISA for selective measurement of adiponectin multimers and identification of adiponectin in human cerebrospinal fluid, *Clin. Chem.* 53 (2007) 1541–1544.
- [23] T.S. Tsao, E. Tomas, H.E. Murrey, C. Hug, D.H. Lee, N.B. Ruderman, et al., Role of disulfide bonds in Acrp30/adiponectin structure and signaling specificity. Different oligomers activate different signal transduction pathways, *J. Biol. Chem.* 278 (2003) 50810–50817.
- [24] U.B. Pajvani, X. Du, T.P. Combs, A.H. Berg, M.W. Rajala, T. Schulthess, et al., Structure-function studies of the adipocyte-secreted hormone Acrp30/adiponectin. Implications for metabolic regulation and bioactivity, *J. Biol. Chem.* 278 (2003) 9073–9085.
- [25] H. Waki, T. Yamauchi, J. Kamon, S. Kita, Y. Ito, Y. Hada, et al., Generation of globular fragment of adiponectin by leukocyte elastase secreted by monocytic cell line THP-1, *Endocrinology* 146 (2005) 790–796.
- [26] Y. Aso, R. Yamamoto, S. Wakabayashi, T. Uchida, K. Takayanagi, K. Takebayashi, et al., Comparison of serum high-molecular weight [HMW] adiponectin with total adiponectin concentrations in type 2 diabetic patients with coronary artery disease using a novel enzyme-linked immunosorbent assay to detect HMW adiponectin, *Diabetes* 55 (2006) 1954–1960.
- [27] Z.V. Wang, T.D. Schraw, J.Y. Kim, T. Khan, M.W. Rajala, A. Follenzi, et al., Secretion of the adipocyte-specific secretory protein adiponectin critically depends on thiol-mediated protein retention, *Mol. Cell. Biol.* 27 (2007) 3716–3731.
- [28] L. Qiang, H. Wang, S.R. Farmer, Adiponectin secretion is regulated by SIRT1 and the endoplasmic reticulum oxidoreductase Ero1-L alpha, *Mol. Cell. Biol.* 27 (2007) 4698–4707.
- [29] M. Liu, L. Zhou, A. Xu, K.S. Lam, M.D. Wetzler, R. Xiang, et al., A disulfide-bond A oxidoreductase-like protein [DsbA-L] regulates adiponectin multimerization, *Proc. Natl. Acad. Sci. U. S. A.* 105 (2008) 18302–18307.
- [30] A.A. Richards, T. Stephens, H.K. Charlton, A. Jones, G.A. Macdonald, J.B. Prins, et al., Adiponectin multimerization is dependent on conserved lysines in the collagenous domain: evidence for regulation of multimerization by alterations in posttranslational modifications, *Mol. Endocrinol.* 20 (2006) 1673–1687.
- [31] C. Caselli, O. Melaiu, M. Maltinti, S. Del Ry, M. Cabiati, T. Prescimone, et al., A methodological reappraisal of total and high molecular weight adiponectin determination in human peripheral circulation: comparison of four immunometric assays, *Clin. Chem. Lab. Med.* 48 (2010) 561–568.
- [32] M. Liu, F. Liu, Transcriptional and post-translational regulation of adiponectin, *Biochem. J.* 425 (2009) 41–52.
- [33] S.A. Phillips, J.T. Kung, Mechanisms of adiponectin regulation and use as a pharmacological target, *Curr. Opin. Pharmacol.* 10 (2010) 676–683.
- [34] N. Maeda, M. Takahashi, T. Funahashi, S. Kihara, H. Nishizawa, K. Kishida, et al., PPARgamma ligands increase expression and plasma concentrations of adiponectin, an adipose-derived protein, *Diabetes* 50 (2001) 2094–2099.
- [35] J. Nakae, Y. Cao, M. Oki, Y. Orba, H. Sawa, H. Kiyonari, et al., Forkhead transcription factor FoxO1 in adipose tissue regulates energy storage and expenditure, *Diabetes* 57 (2008) 563–576.
- [36] L. Qiao, J. Shao, SIRT1 regulates adiponectin gene expression through Foxo1-C/enhancer-binding protein alpha transcriptional complex, *J. Biol. Chem.* 281 (2006) 39915–39924.
- [37] X. Zhang, Z.Z. Li, D.F. Liu, X. Xu, Z.C. Mei, W. Shen, Angiotensin-converting enzyme inhibitors improve hepatic steatosis by modulating expression of tumour necrosis factor-alpha, interleukin-6 and adiponectin receptor-2 in rats with type 2 diabetes, *Clin. Exp. Pharmacol. Physiol.* 36 (2009) 631–636.
- [38] J.Y. Lim, W.H. Kim, S.I. Park, GO6976 prevents TNF-alpha-induced suppression of adiponectin expression in 3T3-L1 adipocytes: putative involvement of protein kinase C, *FEBS Lett.* 582 (2008) 3473–3478.
- [39] K.Y. Kim, J.K. Kim, J.H. Jeon, S.R. Yoon, I. Choi, Y. Yang, c-Jun N-terminal kinase is involved in the suppression of adiponectin expression by TNF-alpha in 3T3-L1 adipocytes, *Biochem. Biophys. Res. Commun.* 327 (2005) 460–467.
- [40] M. Fasshauer, S. Kralisch, M. Klier, U. Lossner, M. Bluher, J. Klein, et al., Adiponectin gene expression and secretion is inhibited by interleukin-6 in 3T3-L1 adipocytes, *Biochem. Biophys. Res. Commun.* 301 (2003) 1045–1050.
- [41] N. Hosogai, A. Fukuhara, K. Oshima, Y. Miyata, S. Tanaka, K. Segawa, et al., Adipose tissue hypoxia in obesity and its impact on adipocytokine dysregulation, *Diabetes* 56 (2007) 901–911.
- [42] K.L. Han, J.S. Choi, J.Y. Lee, J. Song, M.K. Joe, M.H. Jung, et al., Therapeutic potential of peroxisome proliferators-activated receptor-alpha/gamma dual agonist with alleviation of endoplasmic reticulum stress for the treatment of diabetes, *Diabetes* 57 (2008) 737–745.
- [43] T. Yamauchi, J. Kamon, Y. Ito, A. Tsuchida, T. Yokomizo, S. Kita, et al., Cloning of adiponectin receptors that mediate antidiabetic metabolic effects, *Nature* 423 (2003) 762–769.
- [44] T. Kadowaki, T. Yamauchi, Adiponectin and adiponectin receptors, *Endocr. Rev.* 26 (2005) 439–451.
- [45] X. Mao, C.K. Kikani, R.A. Riojas, P. Langlais, L. Wang, F.J. Ramos, et al., APPL1 binds to adiponectin receptors and mediates adiponectin signalling and function, *Nat. Cell Biol.* 8 (2006) 516–523.
- [46] K.K. Cheng, K.S. Lam, Y. Wang, Y. Huang, D. Carling, D. Wu, et al., Adiponectin-induced endothelial nitric oxide synthase activation and nitric oxide production are mediated by APPL1 in endothelial cells, *Diabetes* 56 (2007) 1387–1394.

- [47] L. Zhou, S.S. Deepa, J.C. Etlzer, J. Ryu, X. Mao, Q. Fang, et al., Adiponectin activates AMP-activated protein kinase in muscle cells via APPL1/LKB1-dependent and phospholipase C/Ca2+/Ca2+/calmodulin-dependent protein kinase kinase-dependent pathways, *J. Biol. Chem.* 284 (2009) 22426–22435.
- [48] J.T. Heiker, D. Kosel, A.G. Beck-Sicking, Molecular mechanisms of signal transduction via adiponectin and adiponectin receptors, *Biol. Chem.* 391 (2010) 1005–1018.
- [49] Y. Xu, N. Wang, F. Ling, P. Li, Y. Gao, Receptor for activated C-kinase 1, a novel binding partner of adiponectin receptor 1, *Biochem. Biophys. Res. Commun.* 378 (2009) 95–98.
- [50] J.T. Heiker, C.M. Wottawah, C. Juhl, D. Kosel, K. Mörl, A.G. Beck-Sicking, Protein kinase CK2 interacts with adiponectin receptor 1 and participates in adiponectin signaling, *Cell. Signal.* 21 (2009) 936–942.
- [51] H.K. Charlton, J. Webster, S. Kruger, F. Simpson, A.A. Richards, J.P. Whitehead, ERp46 binds to AdipoR1, but not AdipoR2, and modulates adiponectin signalling, *Biochem. Biophys. Res. Commun.* 392 (2010) 234–239.
- [52] C. Hug, J. Wang, N.S. Ahmad, J.S. Bogan, T.S. Tsao, H.F. Lodish, T-cadherin is a receptor for hexameric and high-molecular-weight forms of Acrp30/adiponectin, *Proc. Natl. Acad. Sci. U. S. A.* 101 (2004 Jul) 10308–10313.
- [53] H. Tanihara, K. Sano, R.L. Heimark, T. St John, S. Suzuki, Cloning of five human cadherins clarifies characteristic features of cadherin extracellular domain and provides further evidence for two structurally different types of cadherin, *Cell Adhes. Commun.* 2 (1994) 15–26.
- [54] T. Takeuchi, Y. Ohtsuki, Recent progress in T-cadherin [CDH13, H-cadherin] research, *Histol. Histopathol.* 16 (2001) 1287–1293.
- [55] T. Takeuchi, Y. Adachi, Y. Ohtsuki, M. Furihata, Adiponectin receptors, with special focus on the role of the third receptor, T-cadherin, in vascular disease, *Med. Mol. Morphol.* 40 (2007) 115–120.
- [56] D. Ivanov, M. Philippova, J. Antropova, F. Gubaeva, O. Iljinskaya, E. Tararak, et al., Expression of cell adhesion molecule T-cadherin in the human vasculature, *Histochem. Cell Biol.* 115 (2001) 231–242.
- [57] M.S. Denzel, M.C. Scimia, P.M. Zumstein, K. Walsh, P. Ruiz-Lozano, B. Ranscht, T-cadherin is critical for adiponectin-mediated cardioprotection in mice, *J. Clin. Invest.* 120 (2010) 4342–4352.
- [58] Y. Takemura, N. Ouchi, R. Shibata, T. Aprahamian, M.T. Kirber, R.S. Summer, et al., Adiponectin modulates inflammatory reactions via calreticulin receptor-dependent clearance of early apoptotic bodies, *J. Clin. Invest.* 117 (2007) 375–386.
- [59] N. Ouchi, J.L. Parker, J.J. Lugus, K. Walsh, Adipokines in inflammation and metabolic disease, *Nat. Rev. Immunol.* 11 (2011) 85–97.
- [60] T. Yamauchi, J. Kamon, Y. Minokoshi, et al., Adiponectin stimulates glucose utilization and fatty-acid oxidation by activating AMP-activated protein kinase, *Nat. Med.* 8 (2002) 1288–1295.
- [61] M. Fasshauer, J. Klein, S. Kralisch, et al., Growth hormone is a positive regulator of adiponectin receptor 2 in 3T3-L1 adipocytes, *FEBS Lett.* 558 (2004) 27–32.
- [62] Amita Yadav, Megha A. Kataria, Vandana Saini, Anil Yadav, Role of leptin and adiponectin in insulin resistance, *Clin. Chim. Acta* 417 (2013) 80–84.
- [63] T. Yamauchi, Y. Nio, T. Maki, M. Kobayashi, T. Takazawa, M. Iwabu, et al., Targeted disruption of AdipoR1 and AdipoR2 causes abrogation of adiponectin binding and metabolic actions, *Nat. Med.* 13 (2007) 332–339.
- [64] M. Bjursell, A. Ahnmark, M. Bohlooly-Y, L. William-Olsson, M. Rhedin, X.R. Peng, et al., Opposing effects of adiponectin receptors 1 and 2 on energy metabolism, *Diabetes* 56 (2007) 583–593.
- [65] Y. Liu, M.D. Michael, S. Kash, W.R. Bensch, B.P. Monia, S.F. Murray, et al., Deficiency of adiponectin receptor 2 reduces diet-induced insulin resistance but promotes type 2 diabetes, *Endocrinology* 148 (2007) 683–692.
- [66] T. Yamauchi, J. Kamon, H. Waki, et al., The fat-derived hormone adiponectin reverses insulin resistance associated with both lipatrophy and obesity, *Nat. Med.* 7 (2001) 941–946.
- [67] A.M. Lincoff, K. Wolski, S.J. Nicholls, S.E. Nissen, Pioglitazone and risk of cardiovascular events in patients with type 2 diabetes mellitus: a meta-analysis of randomized trials, *JAMA* 298 (2007) 1180–1188.
- [68] T. Yamauchi, J. Kamon, H. Waki, K. Murakami, K. Motojima, K. Komeda, et al., The mechanisms by which both heterozygous peroxisome proliferator-activated receptor gamma (PPARgamma) deficiency and PPARgamma agonist improve insulin resistance, *J. Biol. Chem.* 276 (2001) 41245–41254.
- [69] T. Yamauchi, J. Kamon, H. Waki, Y. Terauchi, N. Kubota, K. Hara, et al., The fat-derived hormone adiponectin reverses insulin resistance associated with both lipatrophy and obesity, *Nat. Med.* 7 (2001) 941–946.
- [70] N. Kubota, Y. Terauchi, T. Kubota, H. Kumagai, S. Itoh, H. Satoh, et al., Pioglitazone ameliorates insulin resistance and diabetes by both adiponectin-dependent and -independent pathways, *J. Biol. Chem.* 281 (2006) 8748–8755.
- [71] S.A. Phillips, J. Kung, T.P. Ciaraldi, C. Choe, L. Christiansen, S. Mudaliar, et al., Selective regulation of cellular and secreted multimeric adiponectin by antidiabetic therapies in humans, *Am. J. Physiol. Endocrinol. Metab.* 297 (2009) E767–E773.
- [72] Z.V. Wang, P.E. Scherer, Adiponectin, cardiovascular function, and hypertension, *Hypertension* 51 (2008) 8–14.
- [73] M.I. Yilmaz, A. Sonmez, K. Caglar, T. Celik, M. Yenicesu, T. Eyleten, et al., Effect of antihypertensive agents on plasma adiponectin levels in hypertensive patients with metabolic syndrome, *Nephrol. (Carlton)* 12 (2007) 147–153.
- [74] X. Hui, K.S. Lam, P.M. Vanhoutte, A. Xu, Adiponectin and cardiovascular health: an update, *Br. J. Pharmacol.* 165 (2012) 574–590.
- [75] J.M. Tishinsky, D.J. Dyck, L.E. Robinson, Lifestyle factors increasing adiponectin synthesis and secretion, *Vitam. Horm.* 90 (2012) 1–30.
- [76] T. Yamauchi, T. Kadowaki, Adiponectin receptor as a key player in healthy longevity and obesity-related diseases, *Cell Metab.* 17 (2013) 185–196.
- [77] M.L. Narasimhan, M.A. Coca, J. Jin, T. Yamauchi, Y. Ito, T. Kadowaki, et al., Osmotin is a homolog of mammalian adiponectin and controls apoptosis in yeast through a homolog of mammalian adiponectin receptor, *Mol. Cell* 17 (2005) 171–180.
- [78] M. Okada-Iwabu, T. Yamauchi, M. Iwabu, T. Honma, K. Hamagami, K. Matsuda, M. Yamaguchi, H. Tanabe, T. Kimura-Someya, M. Shirouzu, H. Ogata, K. Tokuyama, K. Ueki, T. Nagano, A. Tanaka, S. Yokoyama, T. Kadowaki, A small-molecule AdipoR agonist for type 2 diabetes and short life in obesity, *Nature* 503 (2013) 493–499.
- [79] D. Kosel, J.T. Heiker, C. Juhl, C.M. Wottawah, M. Blüher, K. Mörl, et al., Dimerization of adiponectin receptor 1 is inhibited by adiponectin, *J. Cell Sci.* 123 (2010) 1320–1328.
- [80] F. Almabouada, A. Diaz-Ruiz, Y. Rabanal-Ruiz, J.R. Peinado, R. Vazquez-Martinez, M.M. Malagon, Adiponectin receptors form homomers and heteromers exhibiting distinct ligand binding and intracellular signaling properties, *J. Biol. Chem.* 288 (2013) 3112–3125.